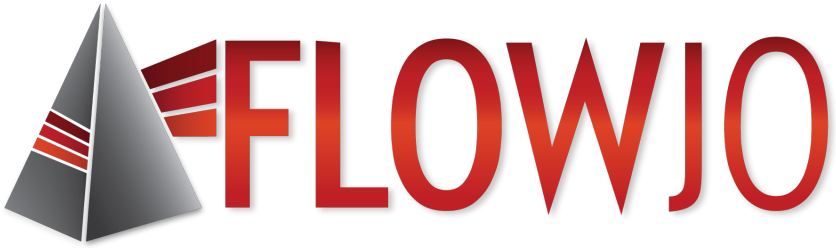
Cytometry Data Analysis in FlowJo V10



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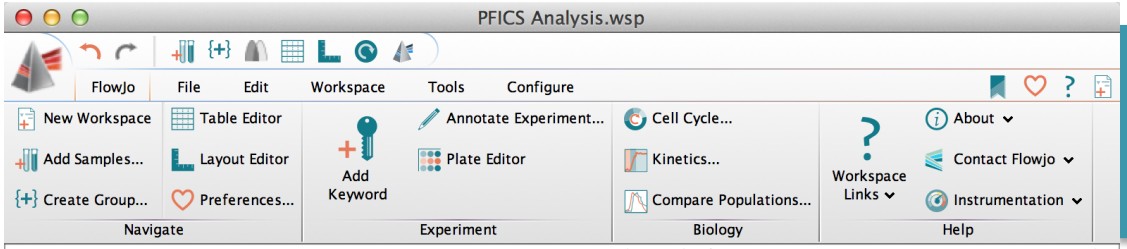
The FlowJo v10 Workspace

- A graphical interface to organize your data.



Ribbon
Tabs and Bands

Group	Size	Role
All Samples	46	Test
All Stain	20	Test
Compensation	12	Compensation
Compensation		
FMOs	14	Controls
Master Gates	34	None
Time		
Singlets		
Lymphocytes		
Live		
CD3+		
Q1: CD4- , CD8+		
Σ Geometric Mean : Comp-Ax488-A (p-ERK1_2)		
IFNg+		
Σ Freq. of Parent		
Perf+		
pERK+		

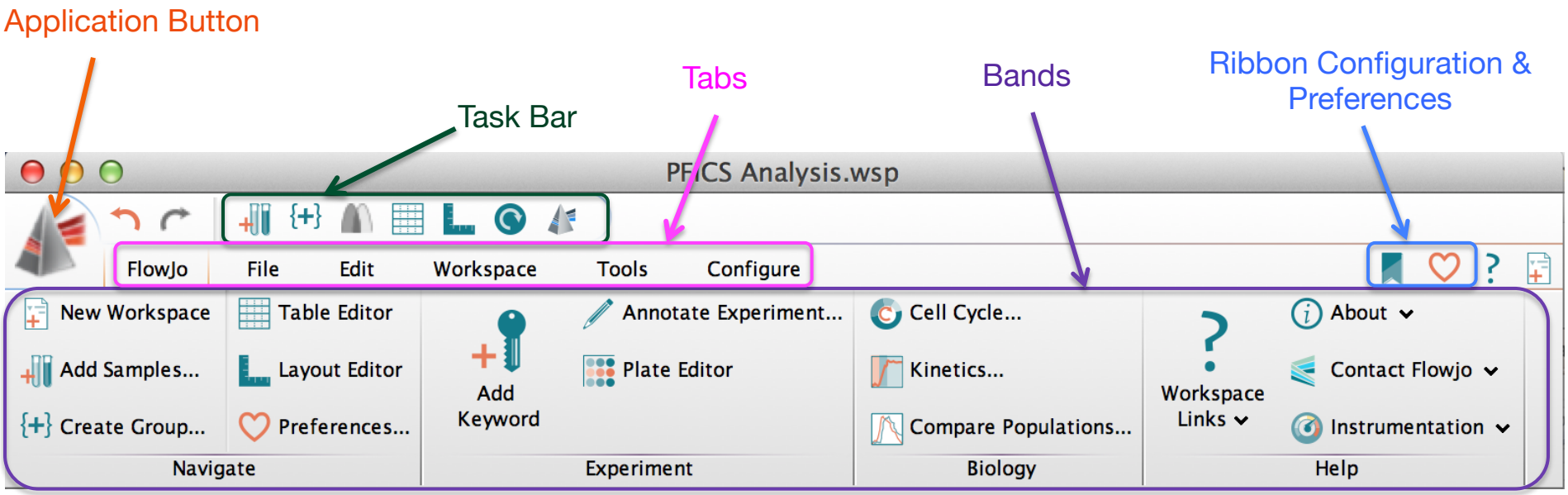
Groups and Group
Analysis

Name	Statistic	#Cells	*PID	*STIM	Well ID:
LD1_NS+NS_A01_exp.fcs		250342	LD1	NS+NS	A01
LD1_NS+PI_C01_exp.fcs		229585	LD1	NS+PI	C01
LD1_PI+NS_B01_exp.fcs		262774	LD1	PI+NS	B01
LD1_PI+PI_D01_exp.fcs		244977	LD1	PI+PI	D01
LD2_NS+NS_A02_exp.fcs		330780	LD2	NS+NS	A02
LD2_NS+PI_C02_exp.fcs		286306	LD2	NS+PI	C02
LD2_PI+NS_B02_exp.fcs		279202	LD2	PI+NS	B02
Time	100.0	279199			
Singlets	96.3	268967			
Lymphocytes	91.3	245663			
Live	73.6	180798			
CD3+	81.7	147761			
Q1: CD4- , CD8+	25.1	37017			
Σ Geometric Mean : Comp-Ax488-A (p-ERK1_2)	424				
IFNg+	64.1	23716			
Σ Freq. of Parent	64.1				
Perf+	52.9	19580			
pERK+	93.2	34514			

Samples and
sample analysis

Ribbons, Tabs and Bands

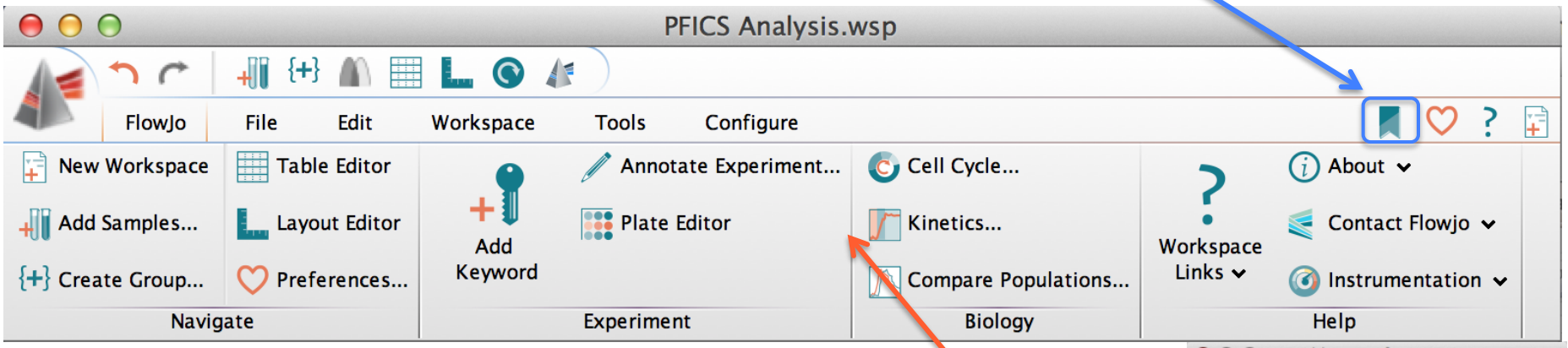
- Ribbon organization allows easy visual navigation of workspace functions.

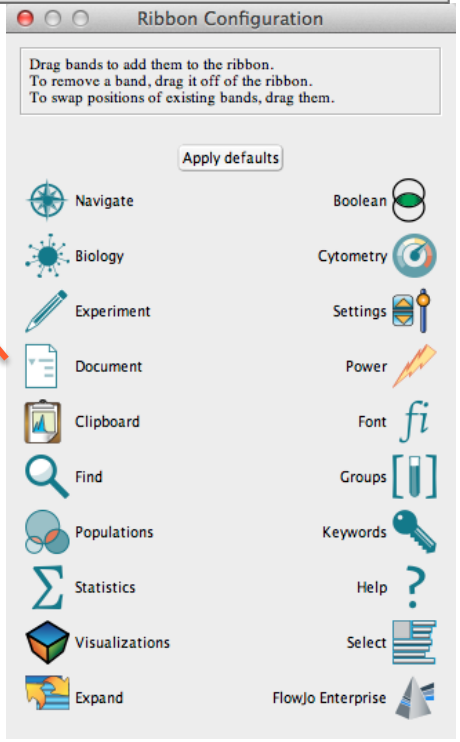


- Tabs group similar Bands together.
- Bands group similar Actions together.

Customizing Ribbons

- Click on the Ribbon icon to configure



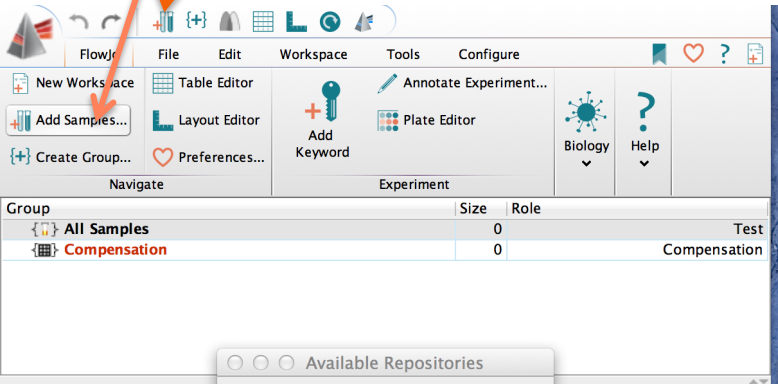


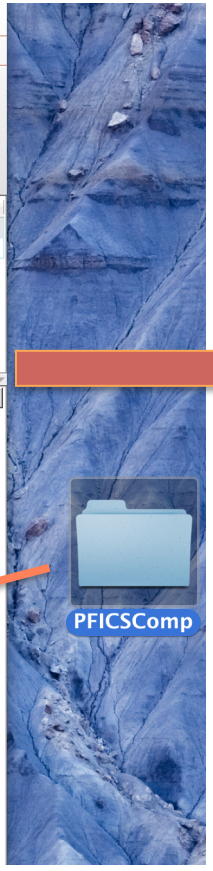
- Drag the icon for any Band into the Ribbon → set of Actions added to your selected Tab.

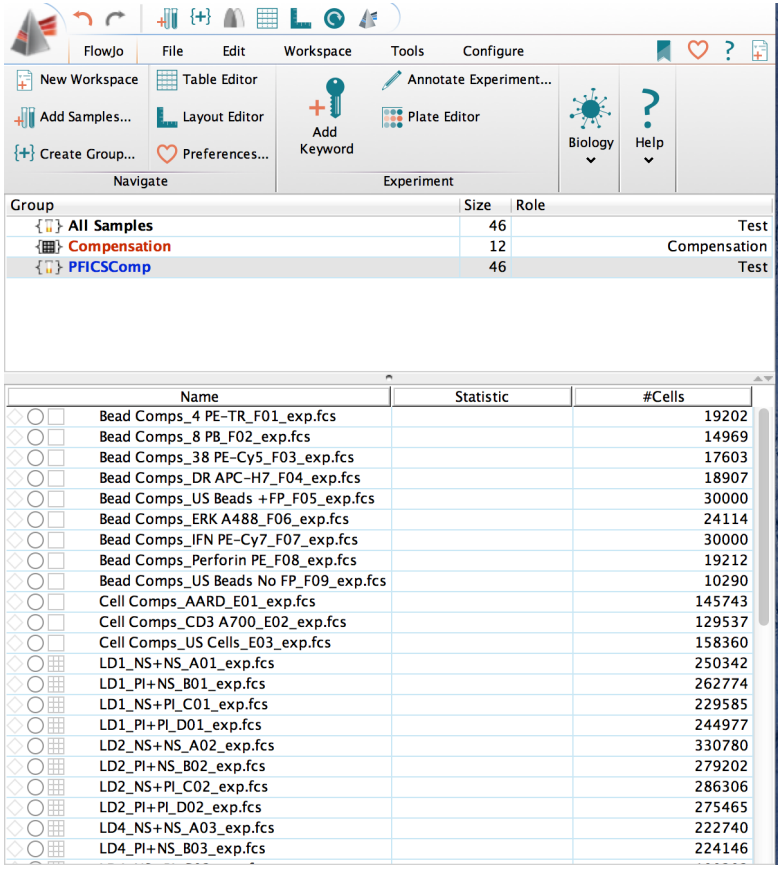
Importing Data

Three possible methods:

1. Drag and drop into samples pane
2. Click Add Samples button
3. Press ⌘ ;







Todays Demo Data Set: Phospho-Flow + Intracellular Cytokine Staining (PFICS)

Polyclonal PFICS Assay:

- Thaw and rest cryopreserved human PBMC overnight
- Stimulate with PMA+Ionomycin (PI) for 2 hours or rest (NS) while blocking protein secretion → signaling and cytokines
- Stain for viability (AARD) and surface antigens (CD3, CD4, CD8, CD38 and HLA-DR)
- Stimulate PI for 20 minutes or NS rest
- Fix, perm and stain for intracellular antigens (phopho-ERK1/2, IFN-γ and Perforin)

PFICS Stim Conditions

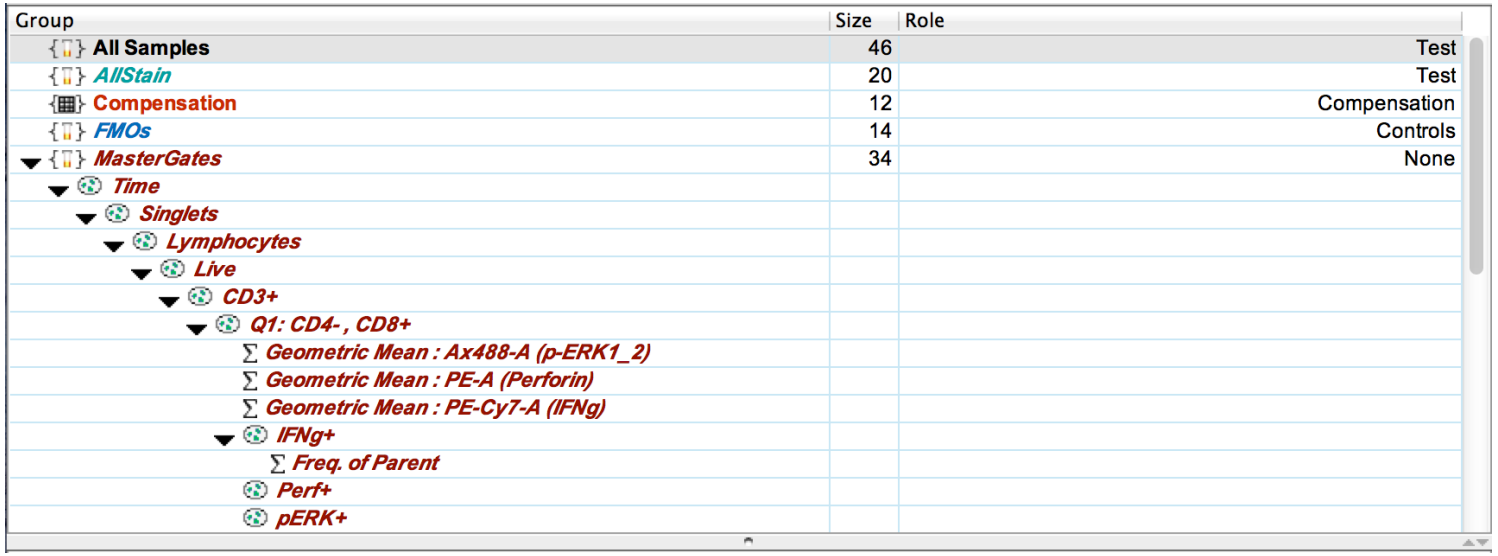
- 2 Stims → 4 potential combinations

Condition	Total Stim Time	phospho-ERK Response	IFN-γ Response
NS+NS	0 min	-	-
NS+PI	20 min	++++	-
PI+NS	120 min	+++	+++
PI+PI	140 min	+++	+++

- 5 donors X 4 stim conditions = 20 experimental All Stain samples
- 1 donor with Fluorescence Minus One (FMO) controls
  7 x 2 stim conditions = 14 FMOs
- 12 Compensation Controls

Group Pane

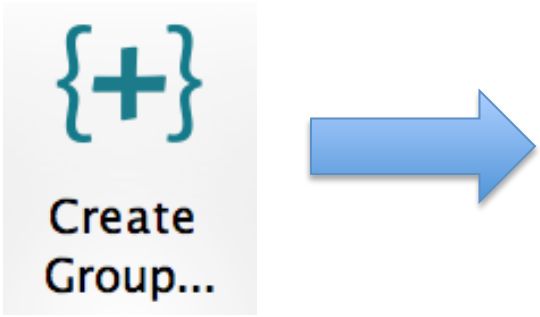
- The Group area lists all groups in the Workspace, # of samples in each group (Size), and the Role of that group (ex. Test, Compensation, Controls) .
- Groups act like folders to organize your samples, allows master gating and unique report generation.

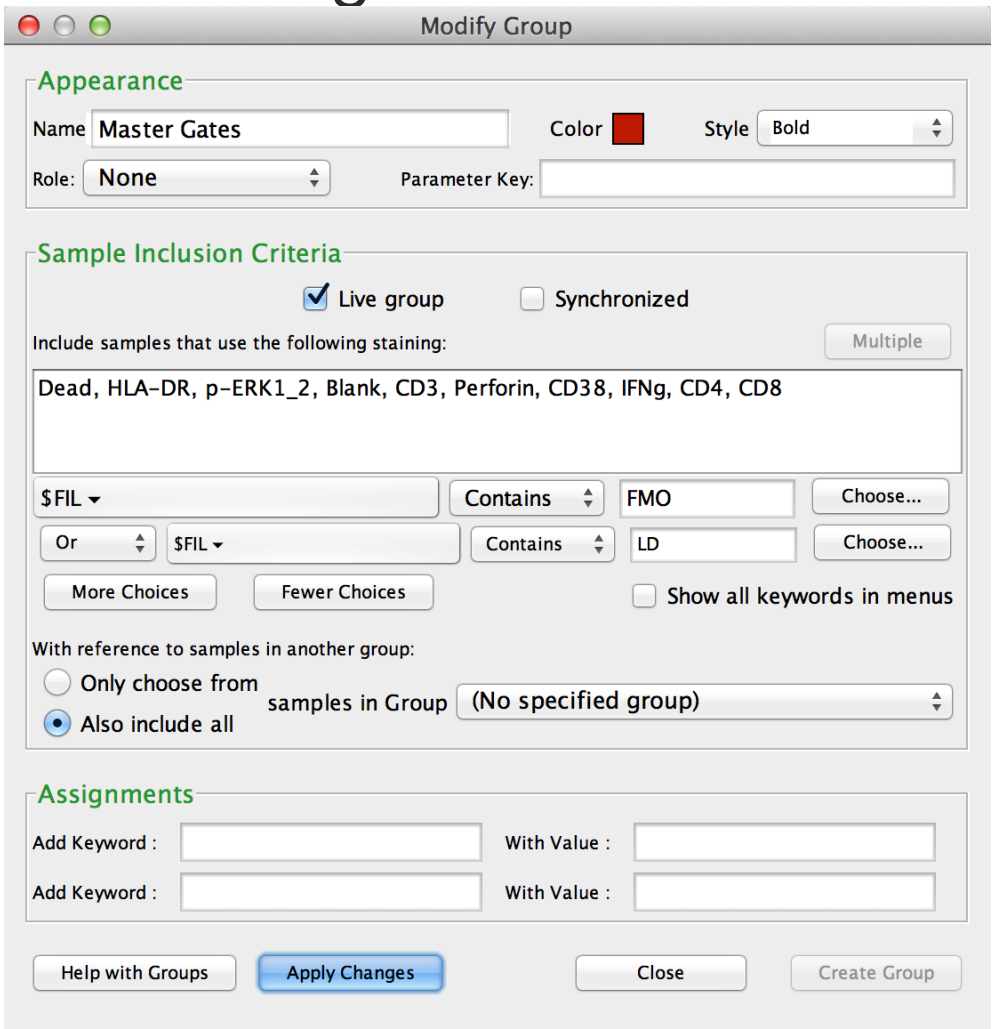


- Group owned analysis gains the group color.

Creating and Editing Groups

- To create a new group type ⌘ G, or click the Create Group Icon located in either the task bar at the top of the workspace, or within the Navigate band.



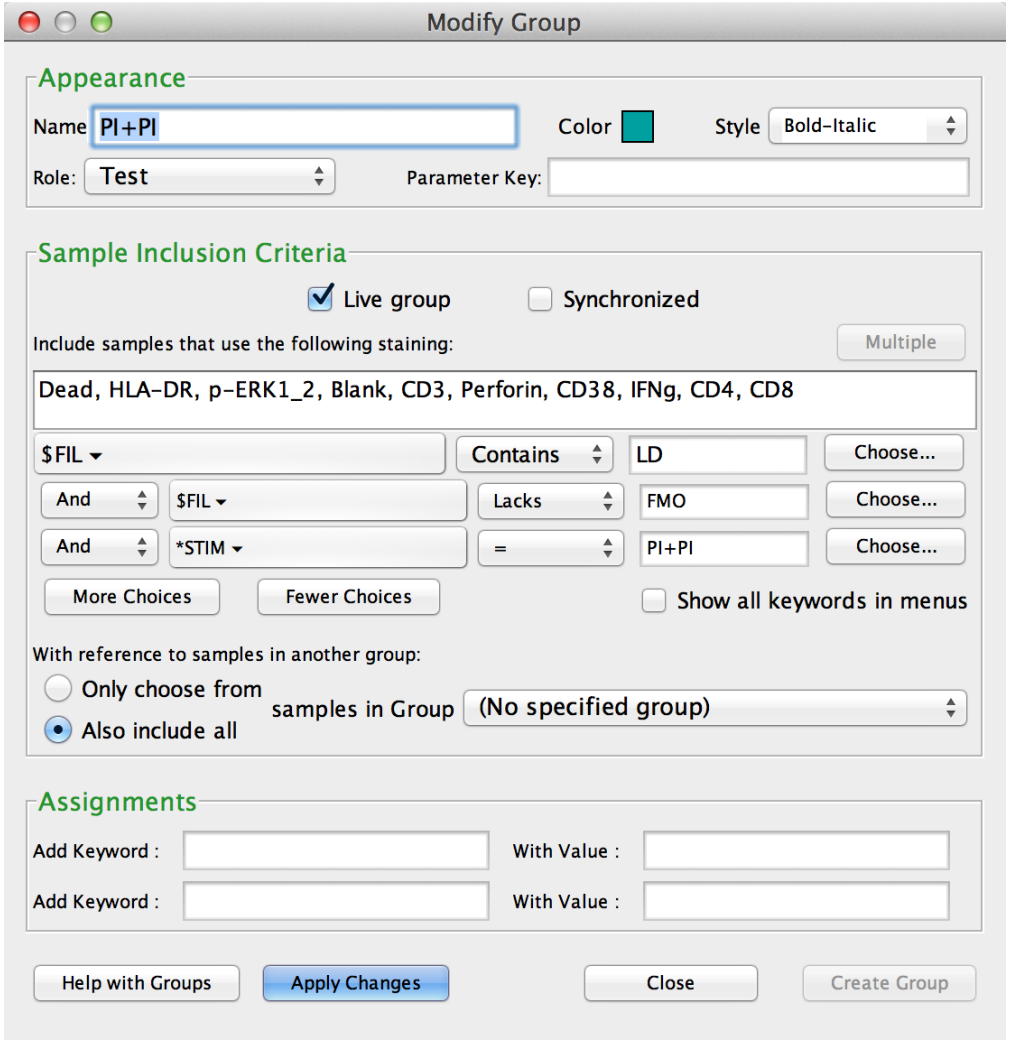


- Double click on an existing group to edit its properties.

Sample Inclusion Criteria

- Live groups automatically include samples based on user-defined inclusion criteria.

- Criteria could include the staining panel, a keyword, characters in the file name, or any combination of these features.



Samples and Sample Analysis

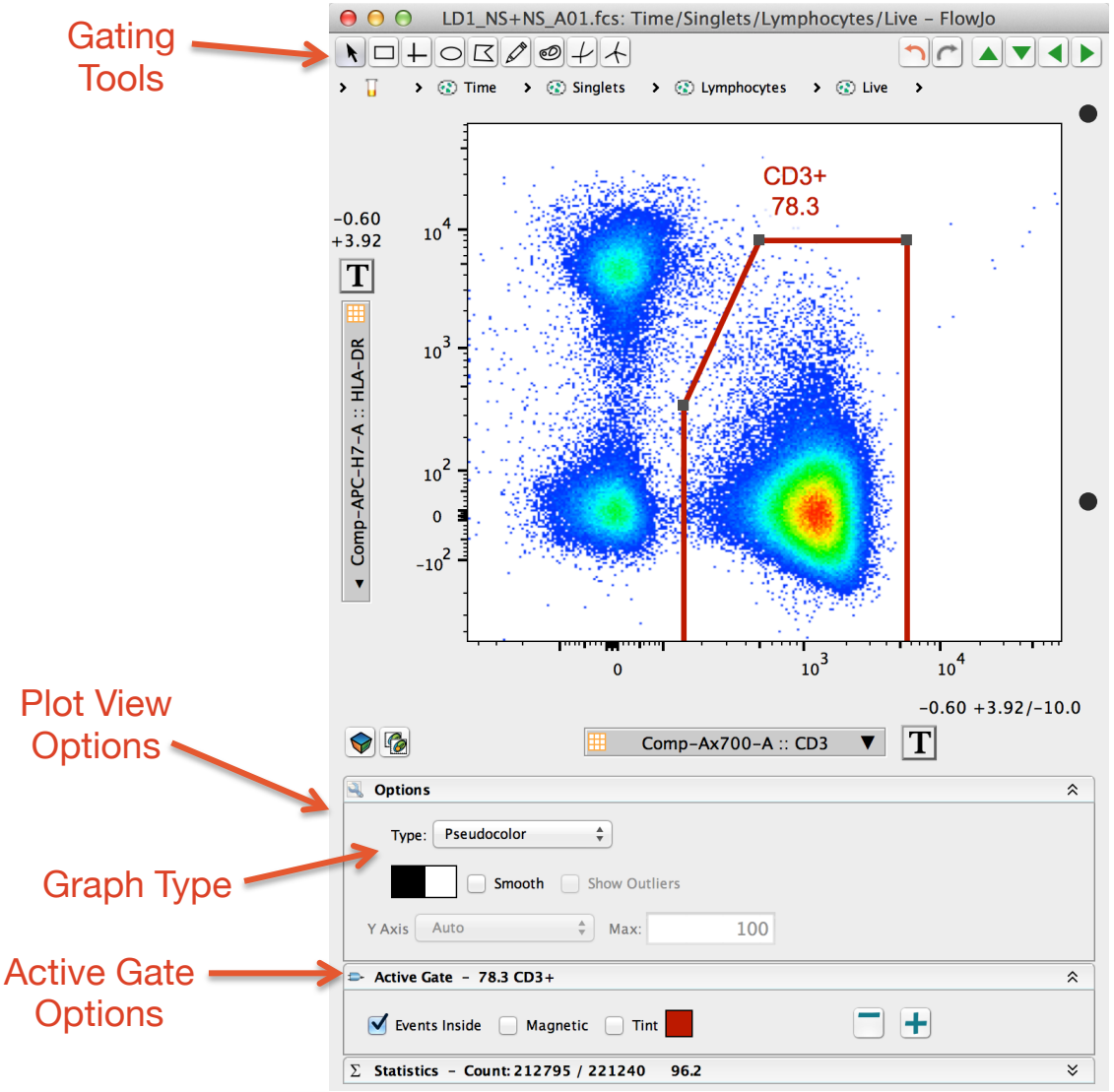
- Displays the sample list and associated analysis of the currently selected group.
- Statistic and #Cells columns are displayed by default. Additional information can be displayed as columns. (Workspace Tab → Add Keywords or Configure Tab → Edit Columns)

Name	Statistic	#Cells	*HIV Status	*PID	*STIM
LD1_NS+NS_A01.fcs		250342	Neg	LD1	NS+NS
LD1_NS+PI_C01.fcs		229585	Neg	LD1	NS+PI
LD1_PI+NS_B01.fcs		262774	Neg	LD1	PI+NS
Time	99.7	261964			
Singlets	96.2	252097			
Lymphocytes	93.7	236200			
Live	96.2	227167			
CD3+	81.4	184893			
Q1: CD4- , CD8+	24.0	44355			
Q2: CD4+ , CD8+	1.13	2090			
Q3: CD4+ , CD8-	72.7	134352			
Q4: CD4- , CD8-	2.22	4096			
LD1_PI+PI_D01.fcs		244977	Neg	LD1	PI+PI

- Double click on a sample to open a Graph Window and add gates.

The Graph Window

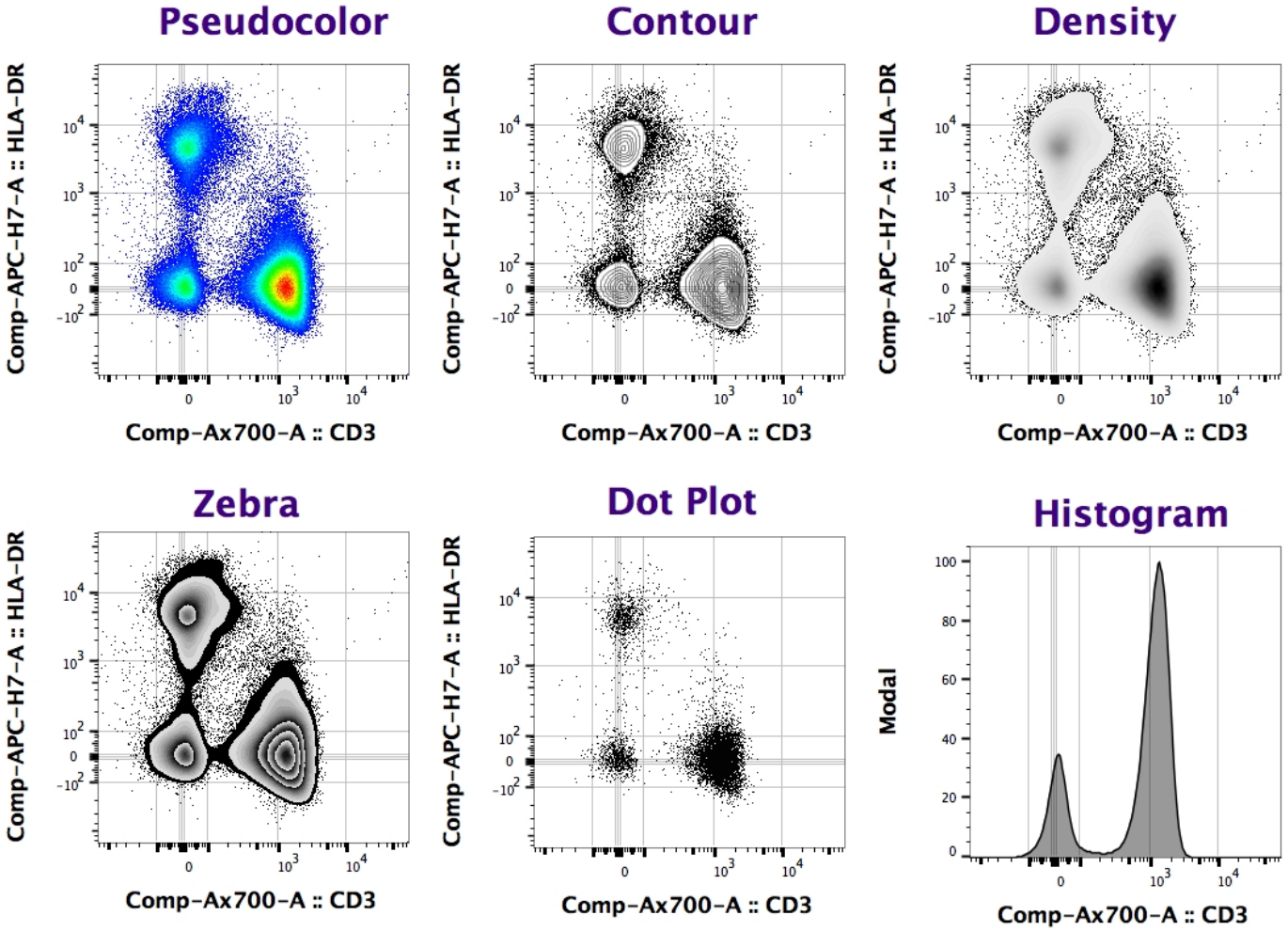
- Facilitates data visualization and gating.



- Several different plot types are available to display flow data.
- Click on the Options Menu below the graph image and select Graph Type from the dropdown menu.

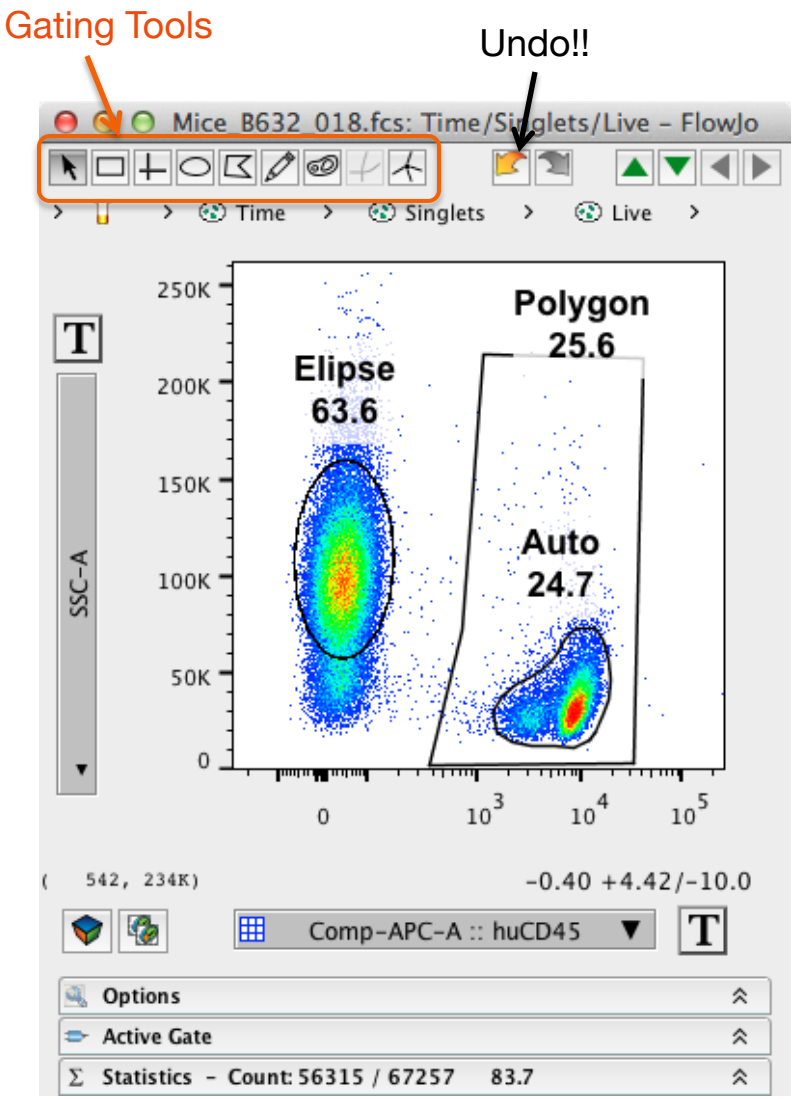
Graph Display Options

- Try them all and pick what pleases you, or best represents your data.



Gating tools

- Are located at the top left in a Graph Window.

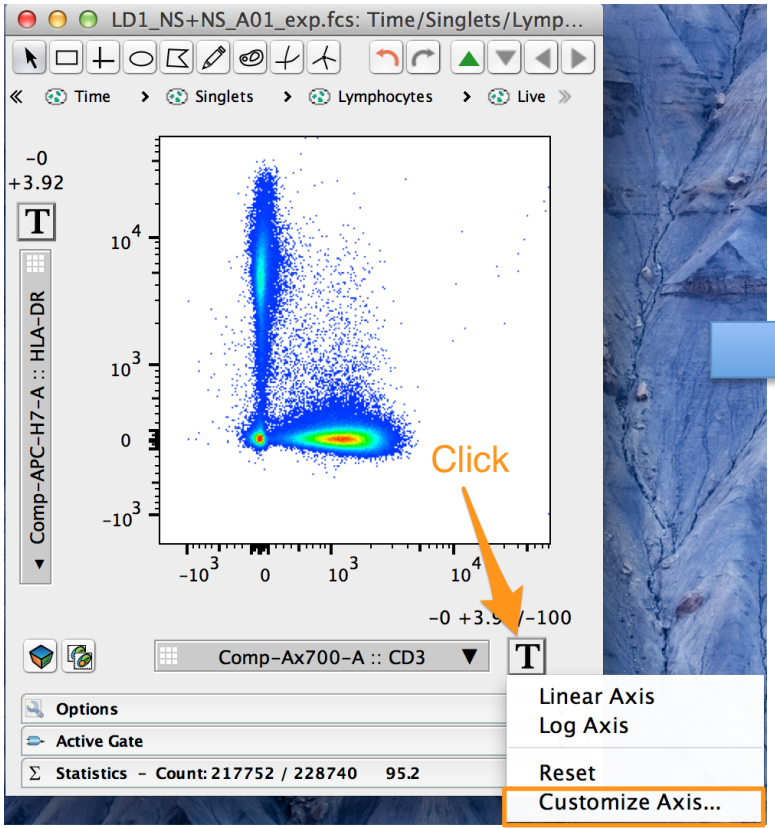


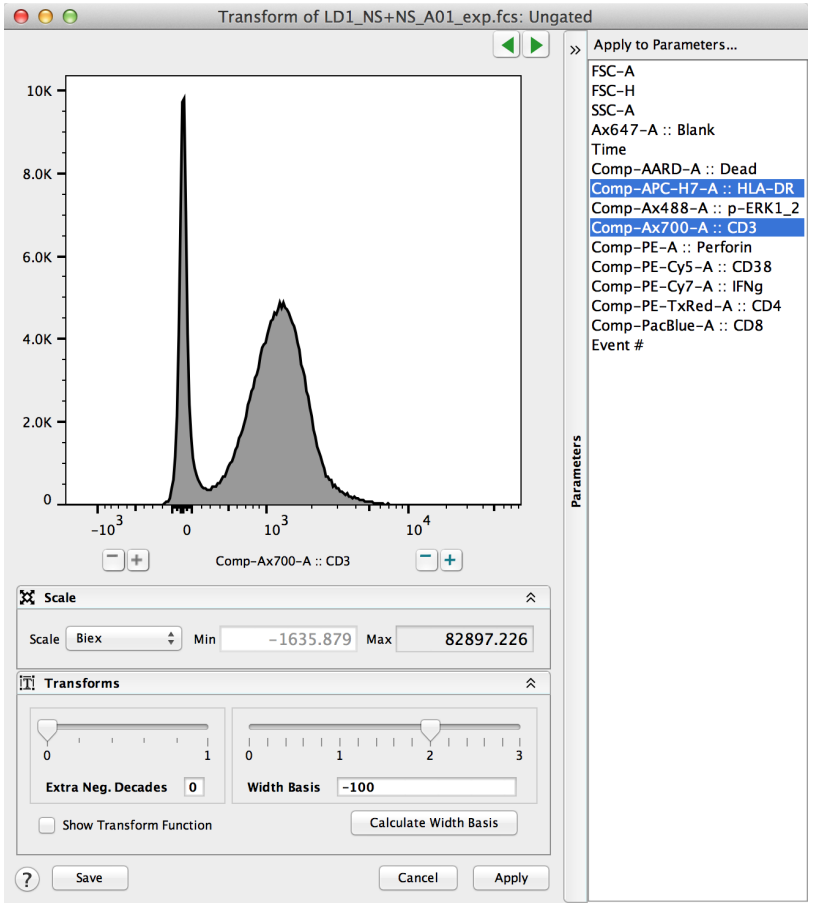
- Gates can always be modified or removed, so don't be shy.

- Explore the gating options and pick what works best for you.

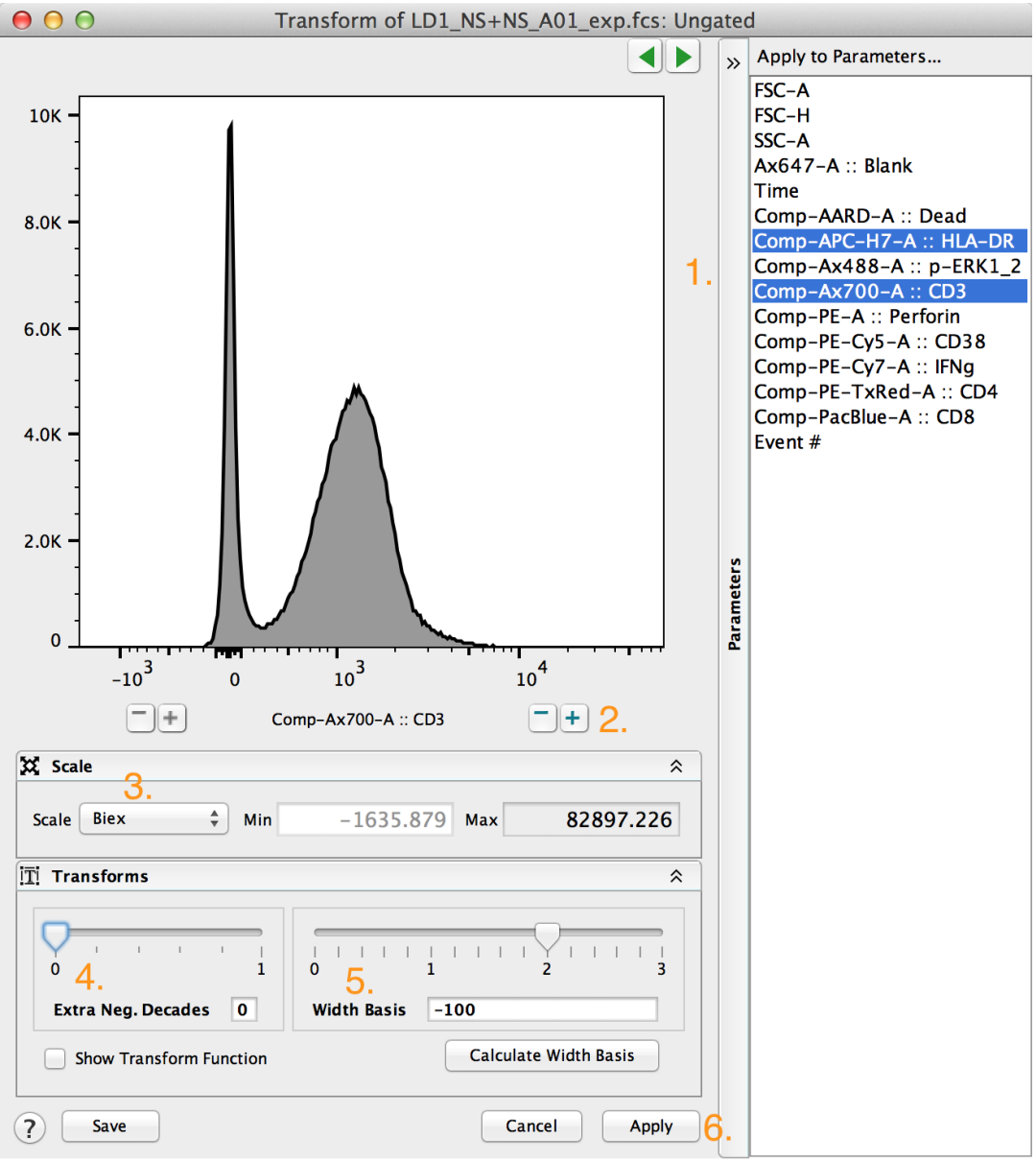
Transforming Data

- Your data may initially look ‘squished’.
- Click the Transformation [T] button and Select Customize Axis… to change the visual display.



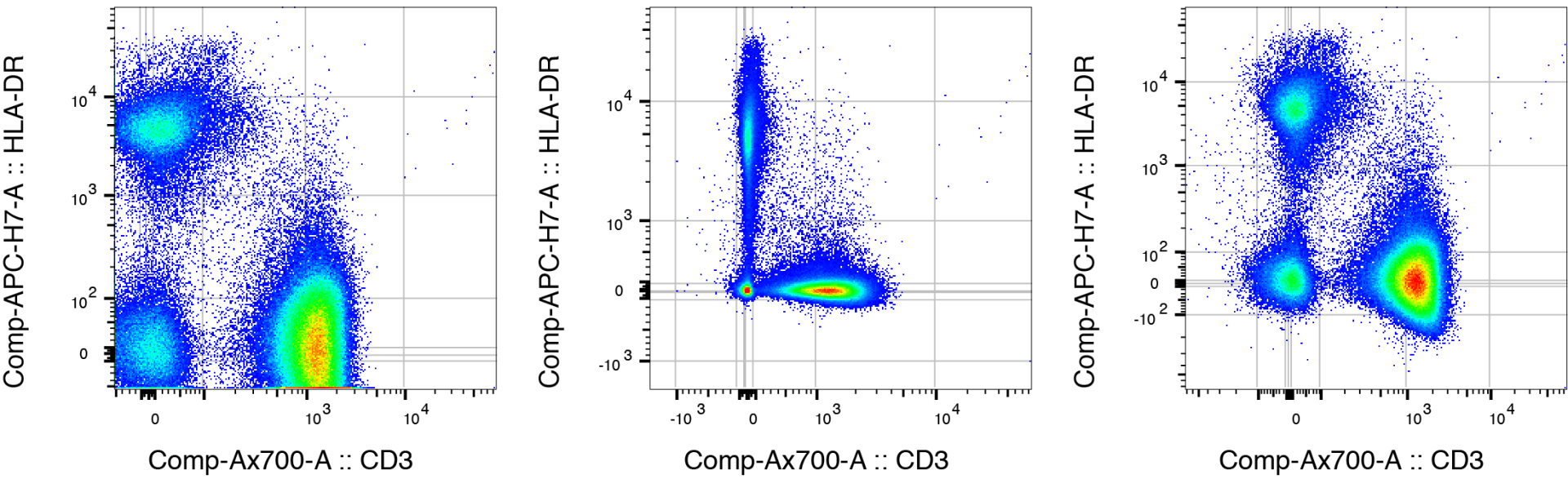


Transform Options



1. Select parameter(s)
2. Add or remove extra Pos. decades/range on top end
3. Select scale (Biex displays linear around zero and log further out)
4. Add or remove extra Neg. decades/range on bottom end
5. Width basis scales how much visual display is given to linear vs. log range of the Biex scale
6. Click the Apply button at bottom right to apply the transformation settings to selected parameters

Effects of Transformation

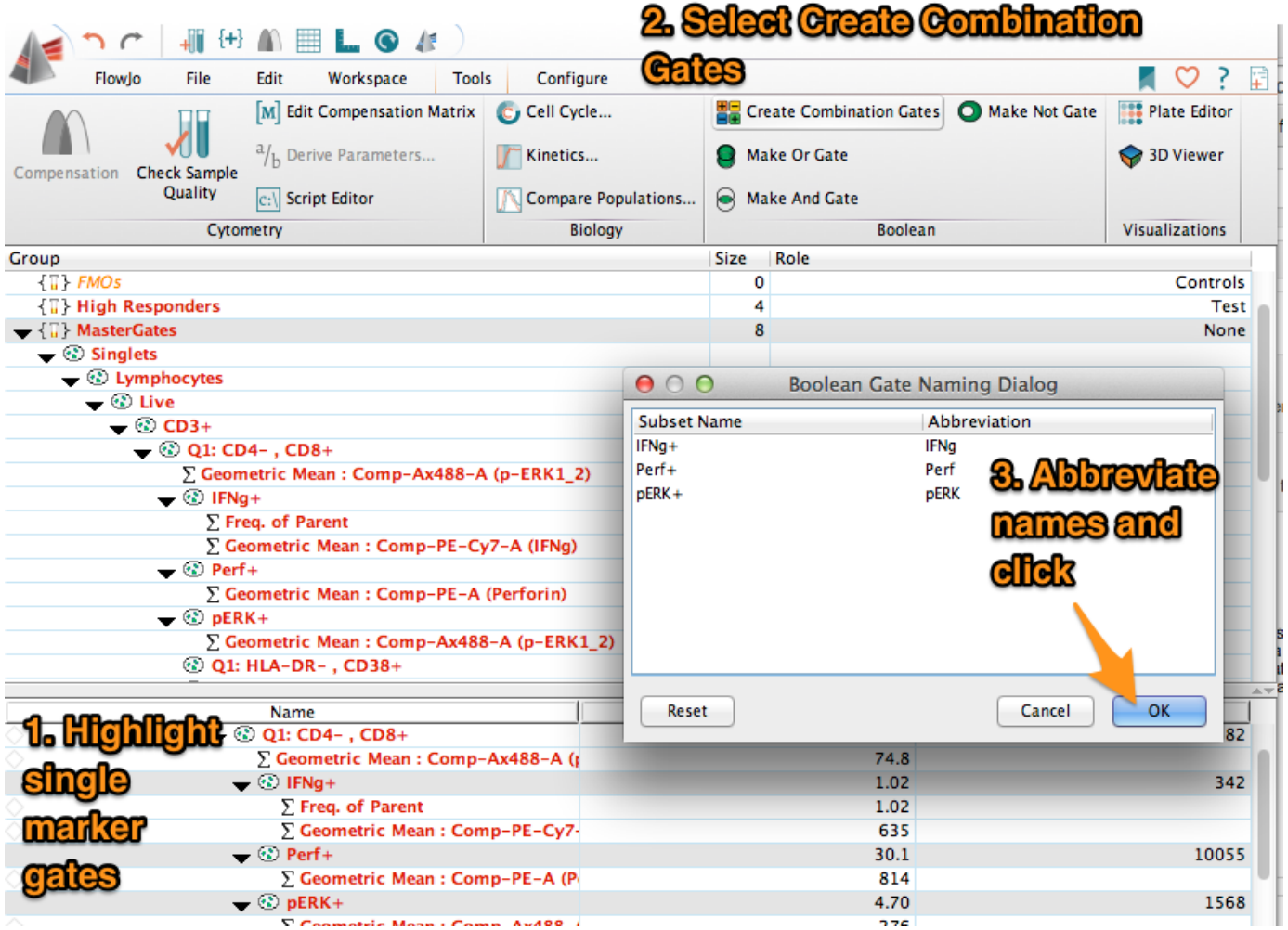


Effects:

1. Gets rid of the “squishing” of cells.
2. Ensures the visual population center better correlates with the statistical center (median).
3. Make high resolution compensated digital cytometry data more appealing to the eye.

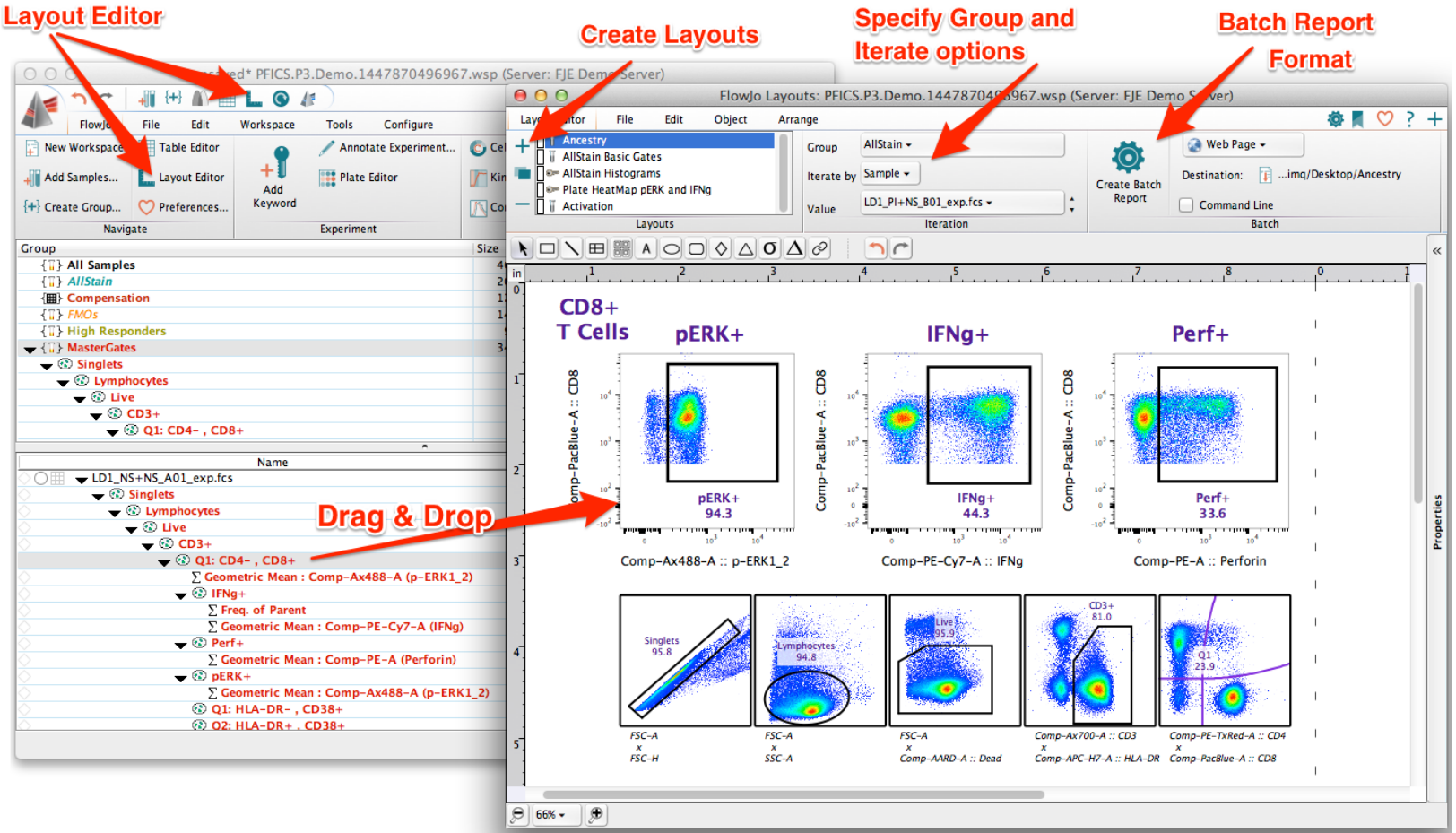
Boolean Combination Gates

- Calculate all possible combinations based on single marker gates (#combinations = $2^{\#gates}$).



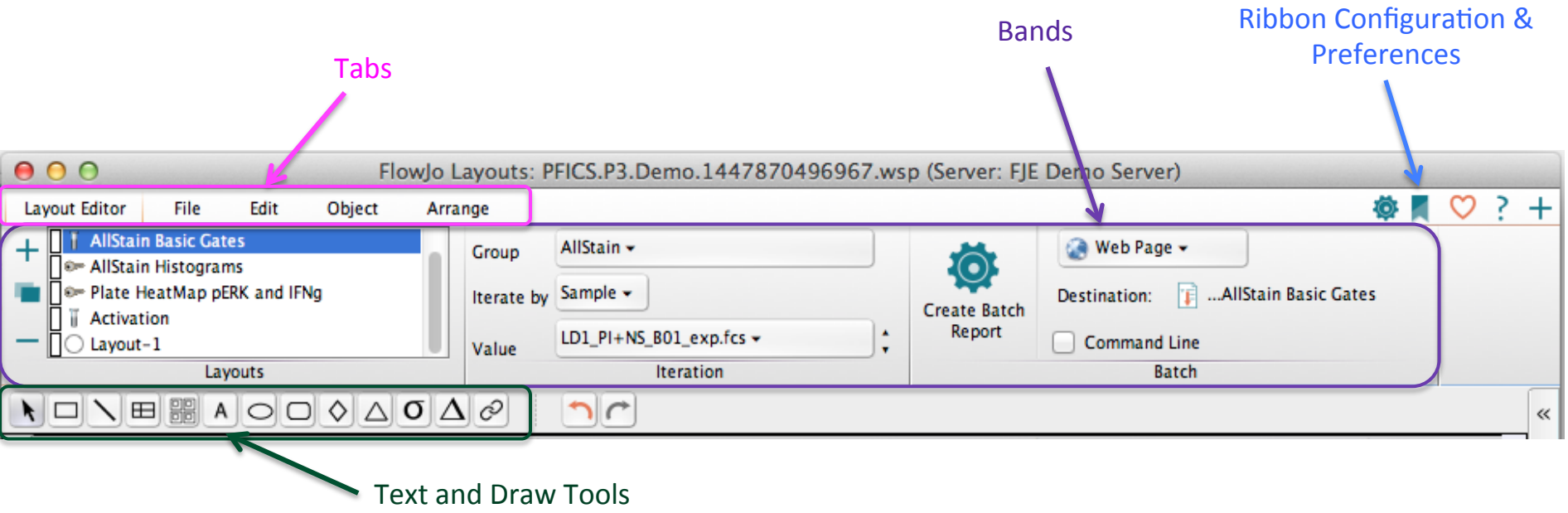
The Layout Editor

- A tool for creating graphical reports.
- Type ⌘ L, or click on the Layout Editor icon.
- Drag populations from a sample to Layout Editor.



Working in Layout Editor

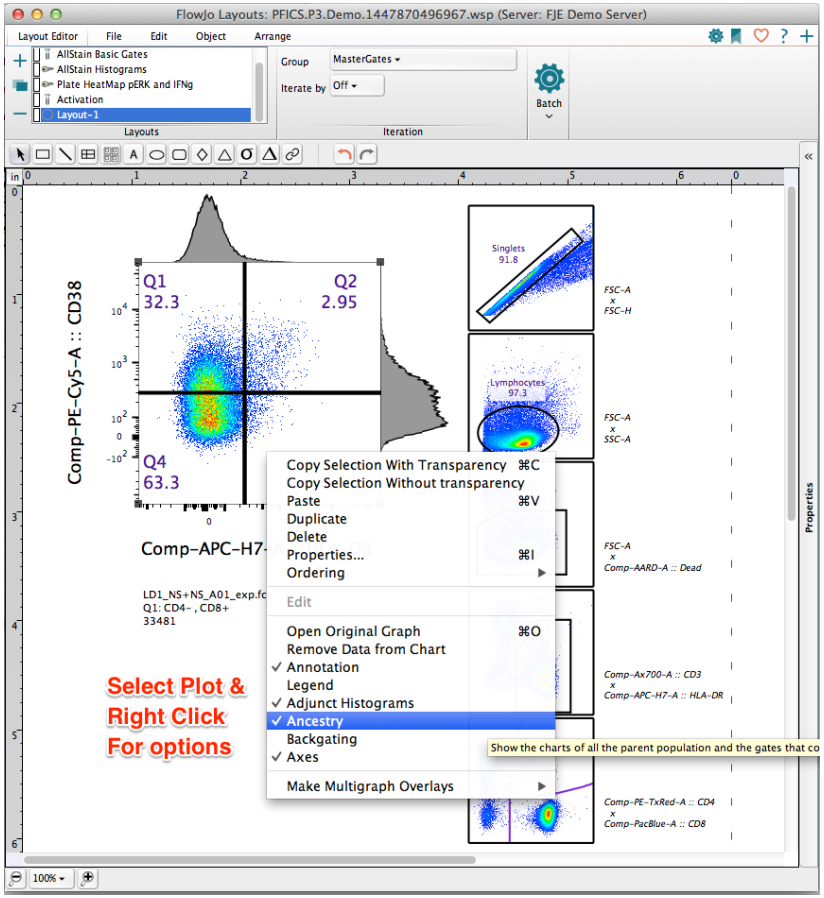
- Similar to the Workspace. Layout Editor has its own customizable Ribbon with Tabs and Bands to organize actions.



- Try clicking on the different tabs to see what types of actions are available.

Within Layout Editor

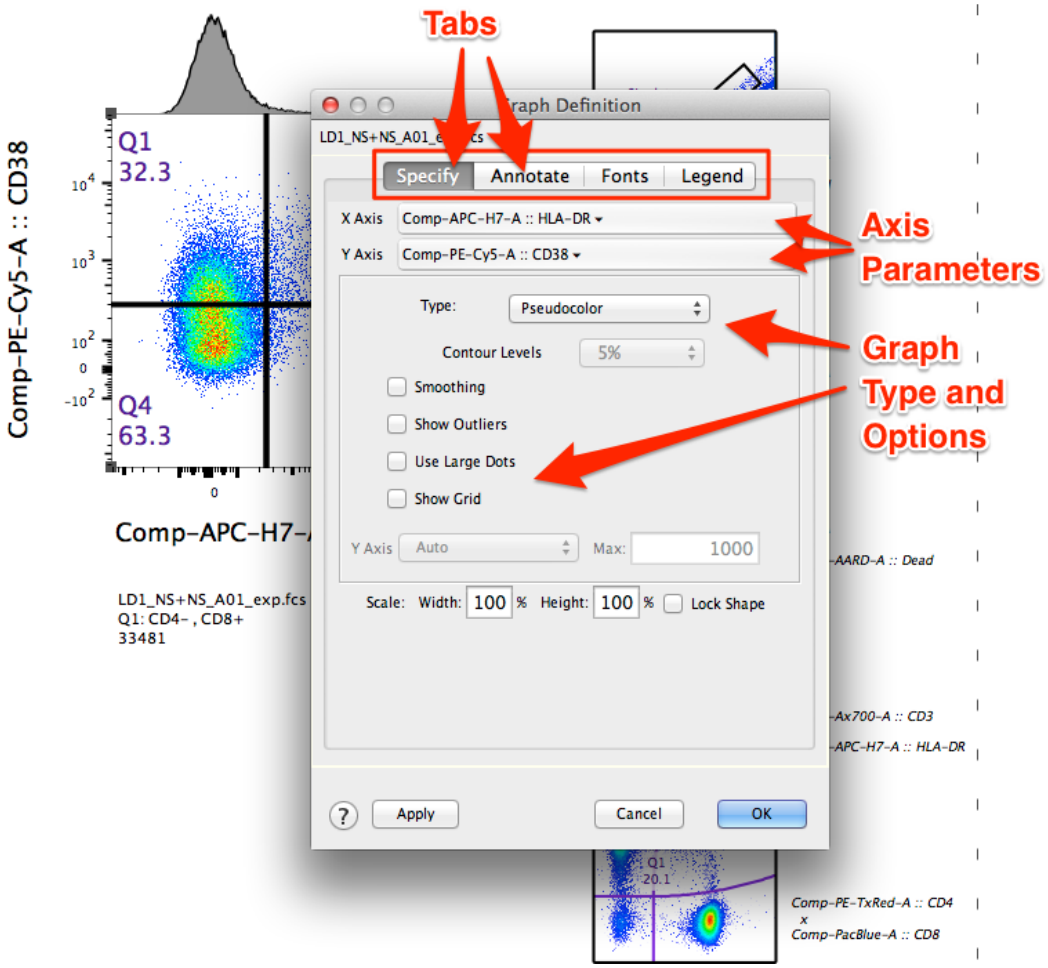
- Graphs can be organized and re-formatted.
- Statistics, keywords, text and even shapes or objects can be added to illustrate your analysis.

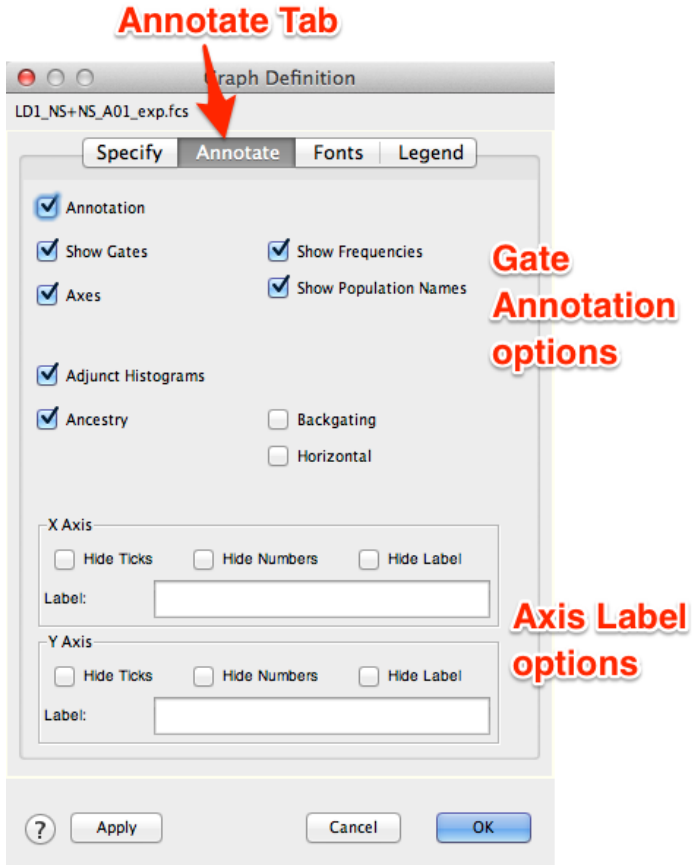


- Right Click on a graph plot for Ancestry and Backgating options
- Right click and select Properties for additional graph formatting

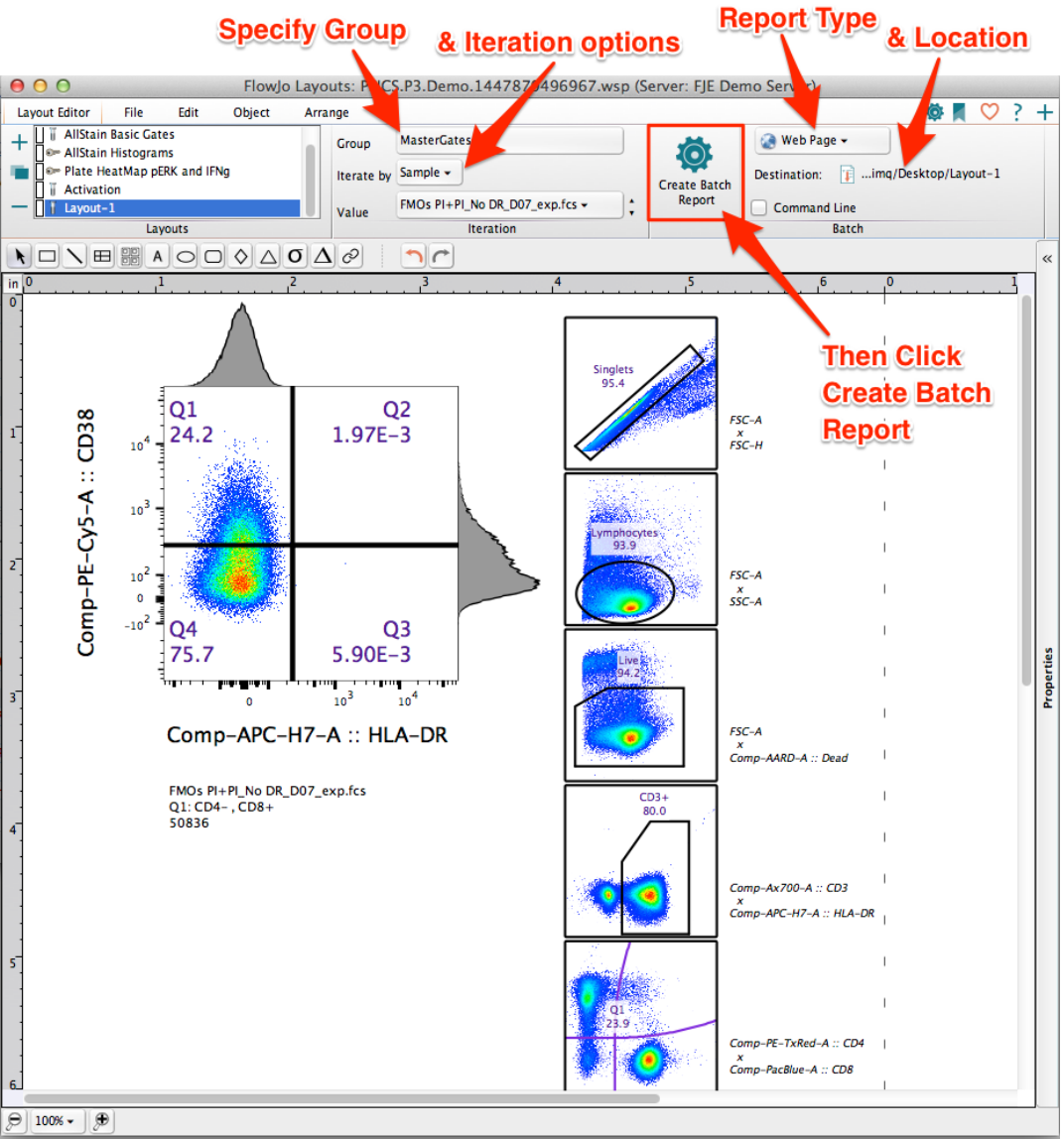
Working in Layout Editor

- Double Click a graph to change its properties/ formatting with 4 tabs of Graph Definition options

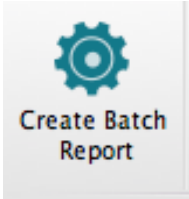




Batch Analysis of Layout Editor Graphics

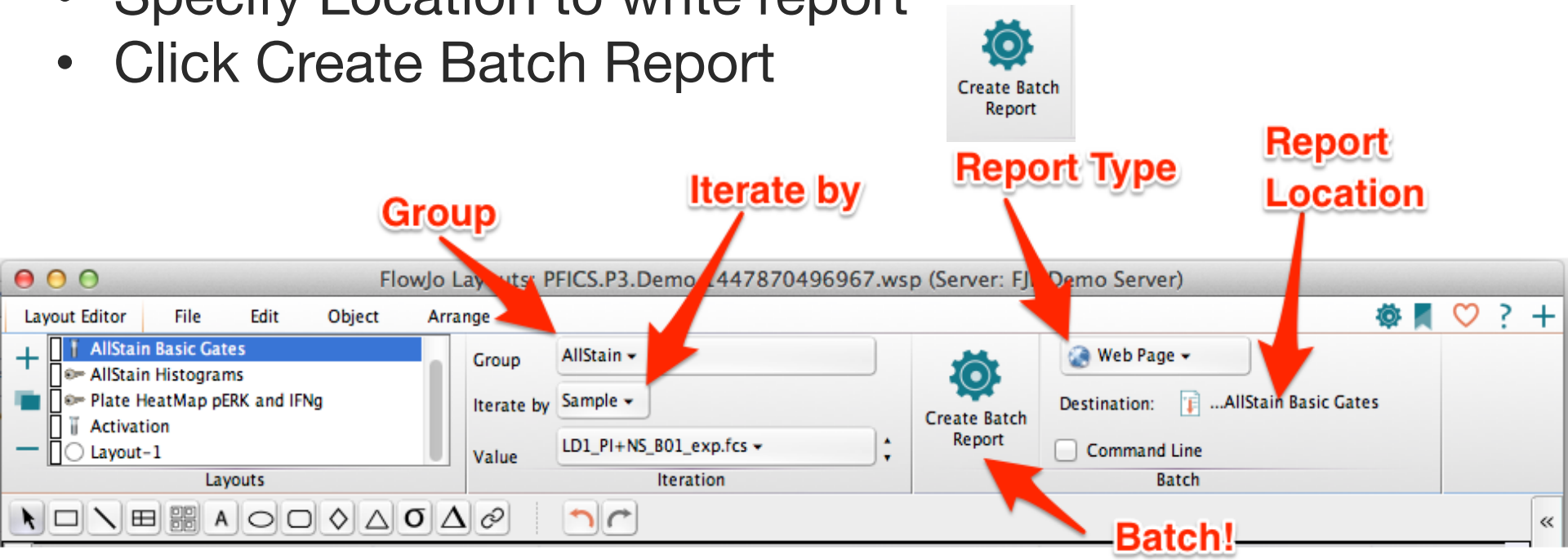


- Batch operations perform repetitive analysis on multiple samples, applying the layout to an entire set of samples.
- Specify Group, Iterate by, Report type and Location, then Click Create Batch Report .



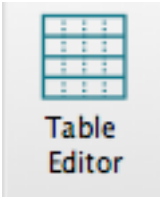
Batch Report Layouts

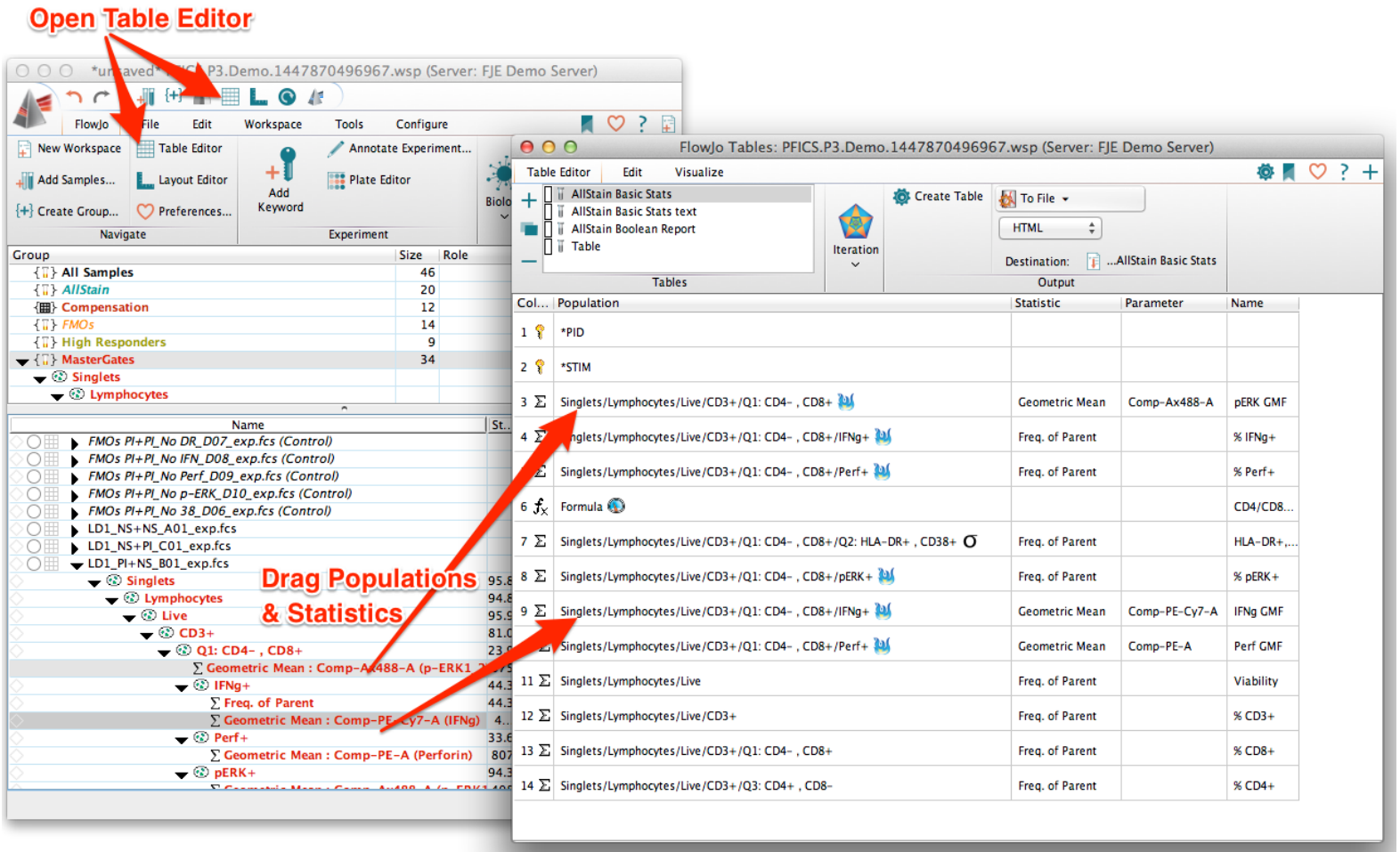
- Specify Group
- Choose Iterate by option
  - Sample
  - Panel
  - Keyword
    - Iterate By (must be Same for all samples displayed in layout)
    - Discriminator (must be Different for all samples displayed in layout)
- Specify type of Report
- Specify Location to write report
- Click Create Batch Report



The Table Editor

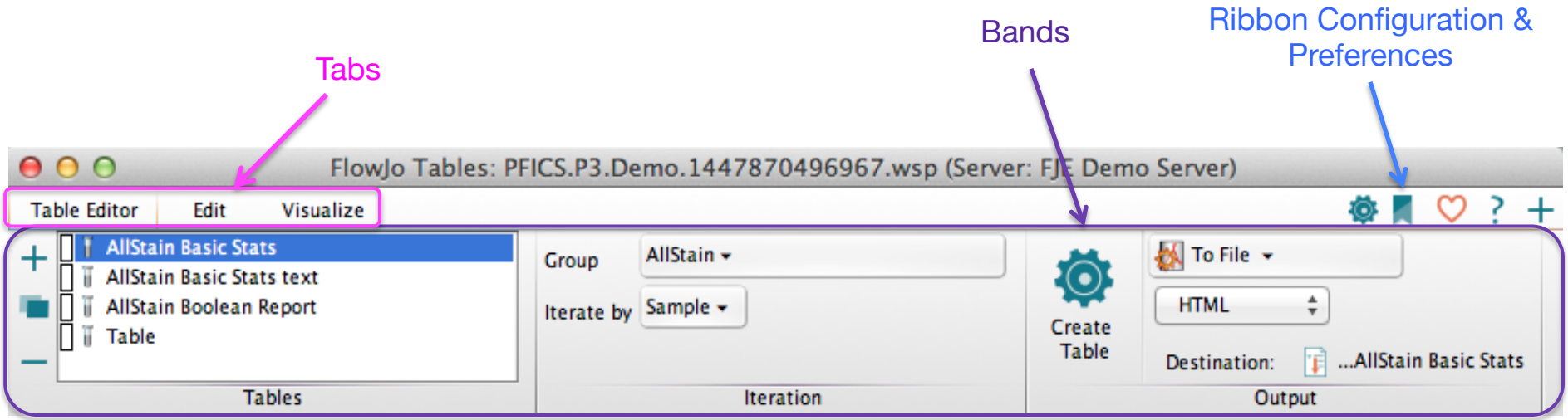
- A tool for creating statistical reports.
- Type ⌘ T, or click on the Table Editor icon.
- Drag Populations & Statistics to Table Editor.





Within Table Editor

- Again, the Table Editor has its own customizable Ribbon with Tabs and Bands to organize actions.

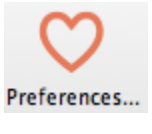


- Specify the group you wish to batch, and how to iterate the batch process, then in the Output band, specify where you want the batch output to go.

Table Editor Visualize Tools

- Table formatting/visualization options such as heat mapping are contained within the Visualize Tab.
- Highlight row(s), then select the visualization.
- Expected Ranges can be set within Preferences

→ Ranges



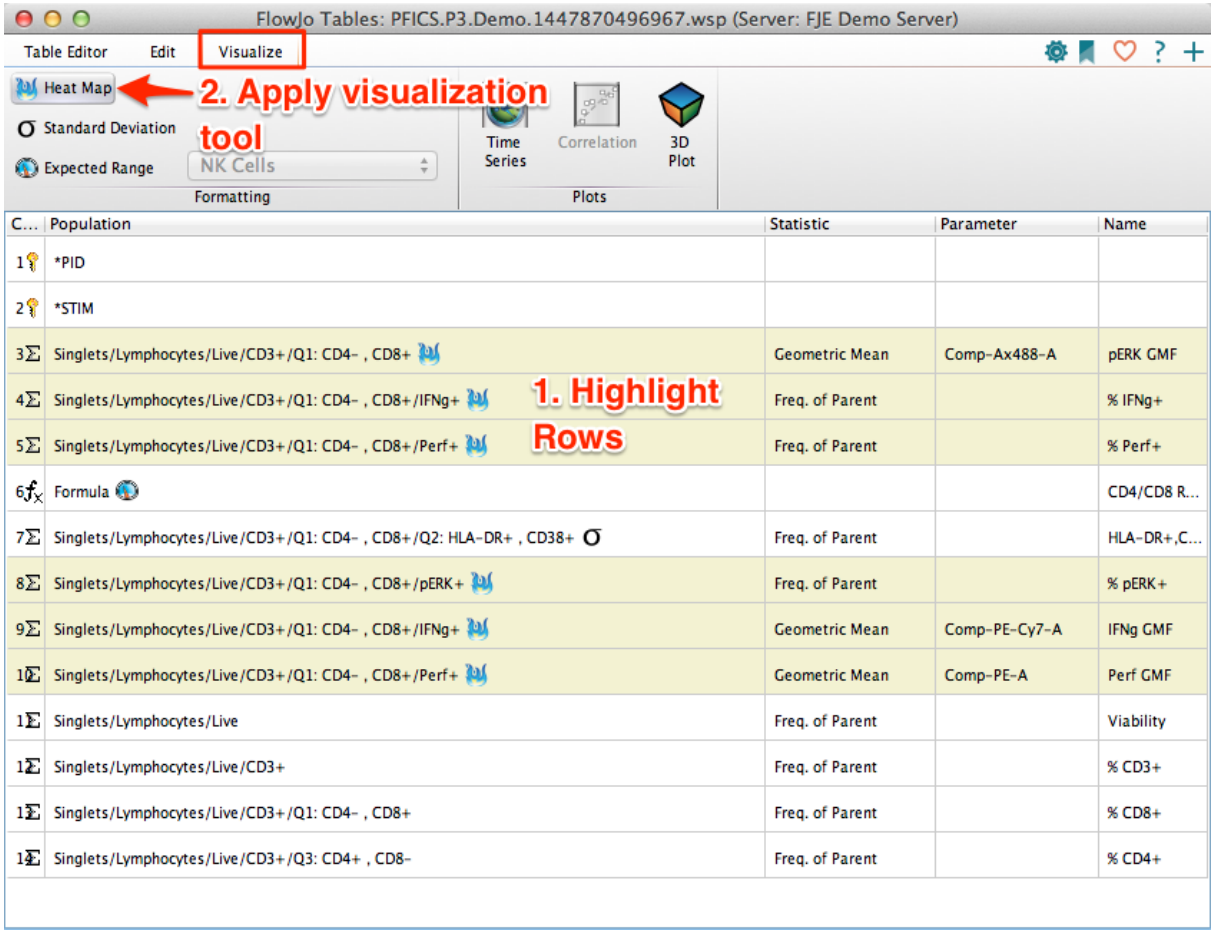


Table Editor Output

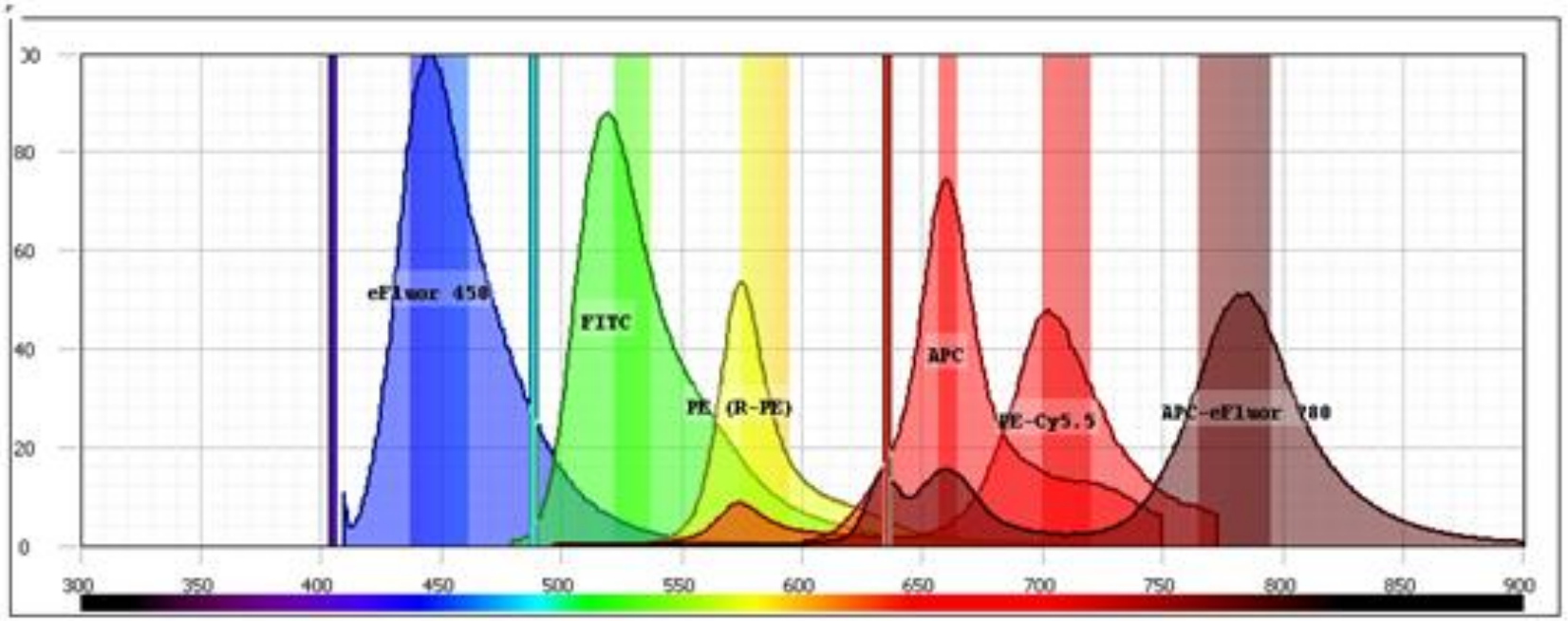
- Formatting/visualization options are maintained when a table is batched to either Display or HTML formats.
- Other file types (ex. Text, CSV, Excel) produce statistics tables lacking visualization formatting.

Table – AllStain Basic Stats

Ancestry Subset Statistic For	*PID	*STIM	pERK GMF	% IFNg+	% Perf+	CD4/CD8 Ratio	HLA-DR+,(	% pERK+	IFNg GMF	Perf GMF
LD1_NS...	LD1	NS+NS	74.1	1.09	30.2	▲ 3.81	*2.95*	4.70	642	812
LD1_NS...	LD1	NS+PI	503	0.96	30.0	▲ 4.13	*2.72*	94.9	504	809
LD1_PI+...	LD1	PI+NS	375	44.3	33.6	▲ 3.04	2.26	94.3	4917	807
LD1_PI+...	LD1	PI+PI	373	43.8	32.7	▲ 3.06	1.94	94.5	4907	816
LD2_NS...	LD2	NS+NS	75.6	1.83	55.9	2.80	2.07	0.45	509	818
LD2_NS...	LD2	NS+PI	496	1.91	53.4	▲ 3.01	1.87	91.0	425	752
LD2_PI+...	LD2	PI+NS	420	64.0	52.1	▲ 2.86	*1.27*	92.6	5894	739
LD2_PI+...	LD2	PI+PI	407	63.7	51.4	▲ 2.91	*1.46*	92.7	5768	734
LD4_NS...	LD4	NS+NS	86.6	1.05	21.1	1.52	*2.71*	8.08	494	740
LD4_NS...	LD4	NS+PI	596	1.74	23.6	1.52	*2.80*	97.1	403	775
LD4_PI+...	LD4	PI+NS	456	28.2	23.8	▼ 1.21	1.74	96.8	5298	577
LD4_PI+...	LD4	PI+PI	449	26.5	22.6	▼ 1.22	*1.48*	96.4	5035	566
LD12_N...	LD12	NS+NS	67.5	0.74	37.5	▲ 3.64	*2.93*	4.14	755	440
LD12_N...	LD12	NS+PI	414	0.50	35.3	▲ 4.28	*3.19*	89.3	683	444
LD12_PI...	LD12	PI+NS	327	45.3	40.8	1.94	*1.50*	84.8	4632	408
LD12_PI...	LD12	PI+PI	319	46.1	41.4	1.94	*1.64*	83.7	4793	403
LD14_N...	LD14	NS+NS	72.4	0.50	14.3	2.11	1.90	4.11	689	811
LD14_N...	LD14	NS+PI	483	0.45	13.8	2.30	2.19	95.5	595	829
LD14_PI...	LD14	PI+NS	366	17.7	18.2	1.66	*1.21*	94.8	3708	650
LD14_PI...	LD14	PI+PI	351	17.0	18.3	1.67	*1.10*	93.2	3565	644
Mean			336	20.4	32.5	2.53	2.05	70.7	2711	679
SD			167	23.0	13.4	0.96	0.65	39.5	2259	152

Compensation

- Compensation corrects for spillover between fluorochrome emission spectra.



- Compensation is essential for multicolor panels

Three Rules of Compensation

- First, there must be a single stained control for every parameter in the experiment!

- In Addition, there are three *rules* for ‘good’ compensation controls.

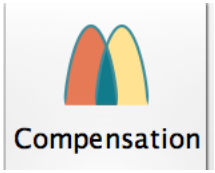
1. Controls need to be at least as bright or brighter than any sample the compensation will be applied to.
2. Background fluorescence should be the same for the positive and negative control.
3. Compensation controls MUST match the exact experimental fluorochrome.

PFICS Compensation Controls

- PBMC Cells
  1. Unstained Cells
  2. AARD
  3. CD3 Alexa700

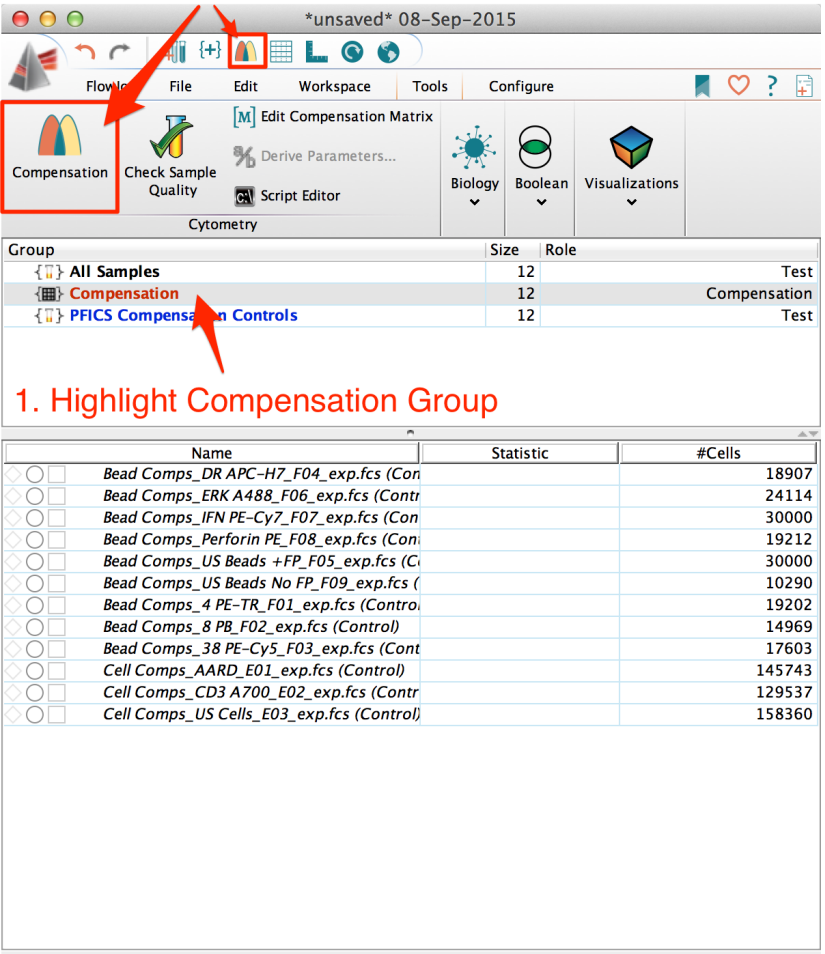
- Compensation Beads
  1. Unstained Beads with Fix and Perm
  2. CD4 PE-TexasRed
  3. CD8 Pacific Blue
  4. CD38 PE-Cy5
  5. HLA-DR APC-H7
  6. Unstained Beads without Fix and Perm
  7. p-ERK1/2 Alexa 488
  8. IFN-g PE-Cy7
  9. Perforin PE

Compensation

- Select a Compensation Group in the groups window, then click 

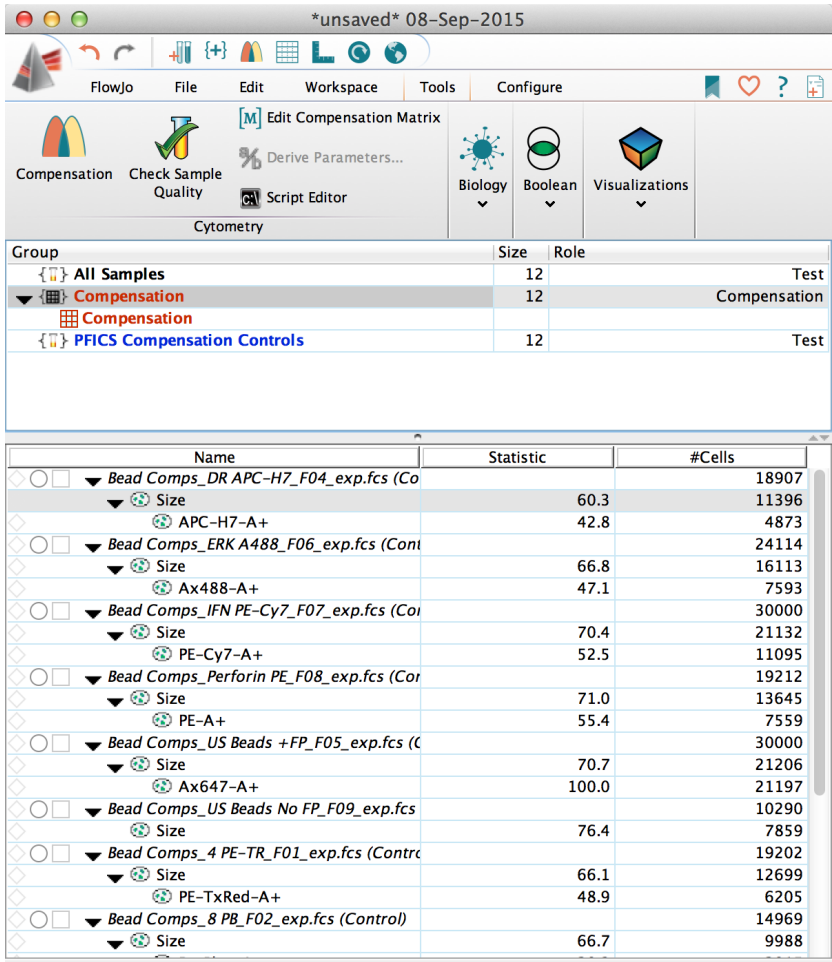
in the task bar.

2. Click the Compensation Tool



1. Highlight Compensation Group

The wizard auto gates samples

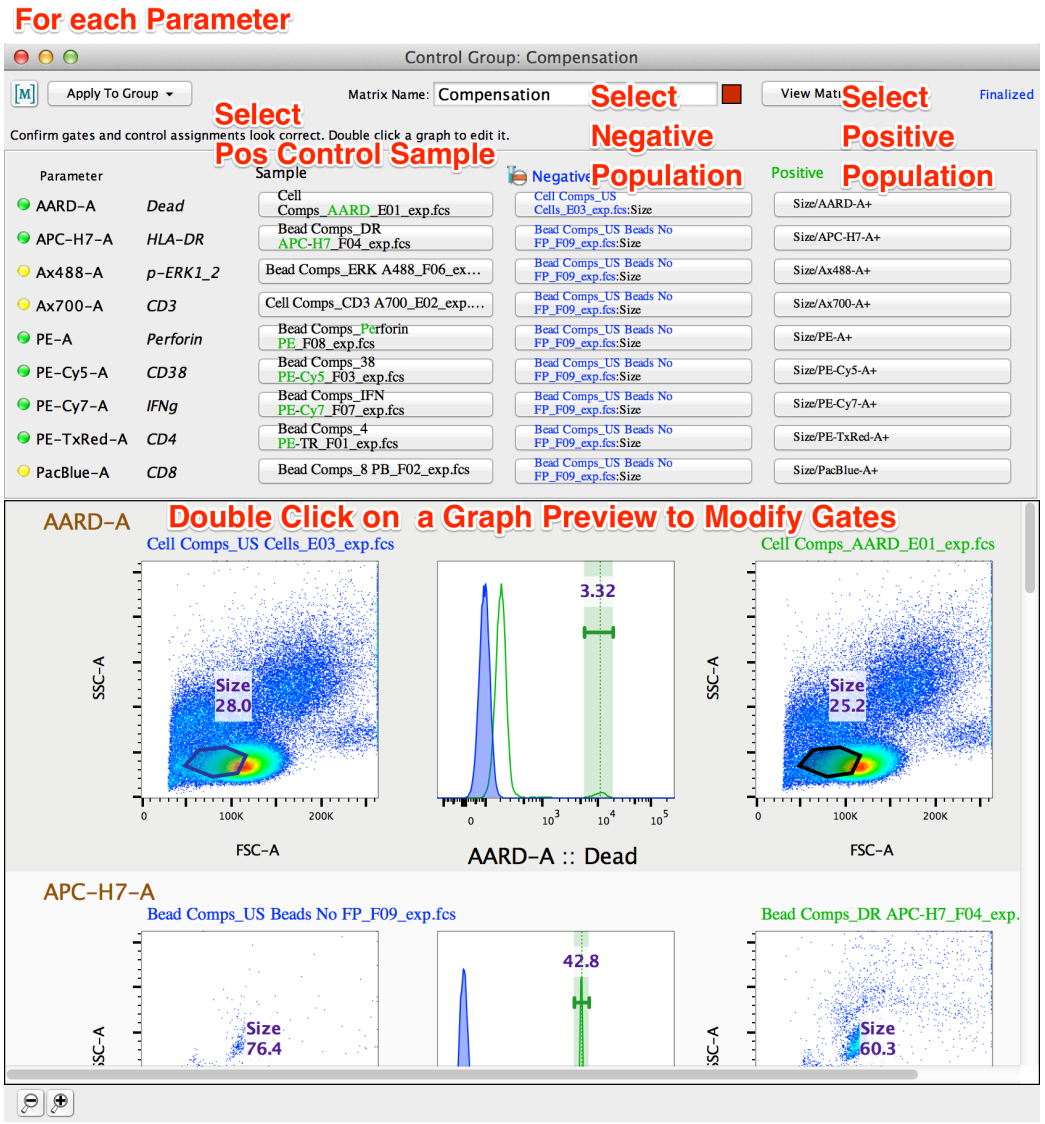


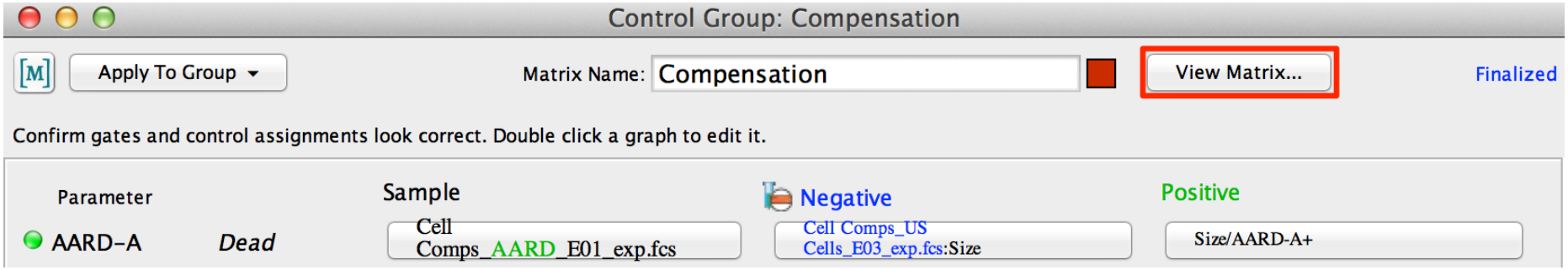
Compensation

- Then fills in the positive and negative.

- Choose from the dropdown lists for each parameter.

- Double click preview graphs to modify gates.

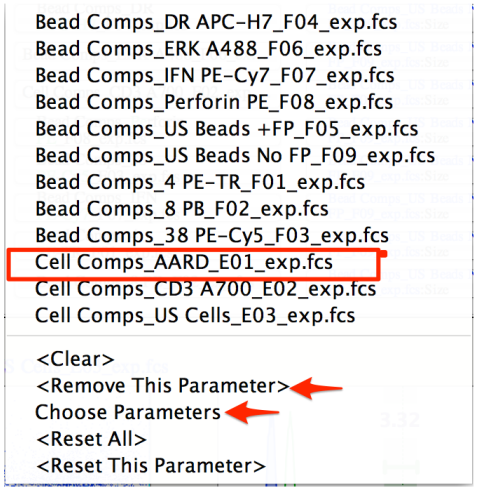




Use Sample drop down list

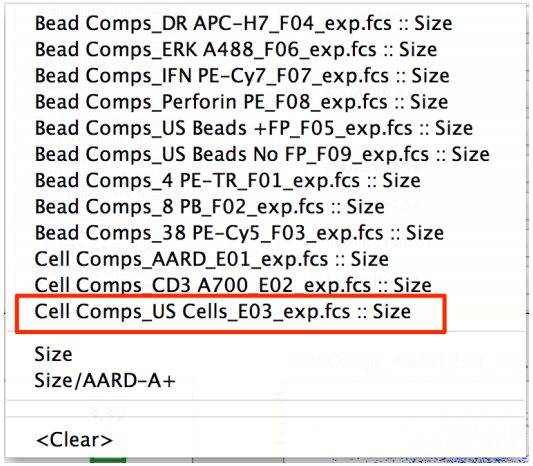
to select Pos Control Sample and

Choose or Remove Parameters



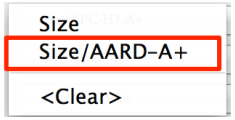
Use Negative drop down list

to Select Negative Sample or Population



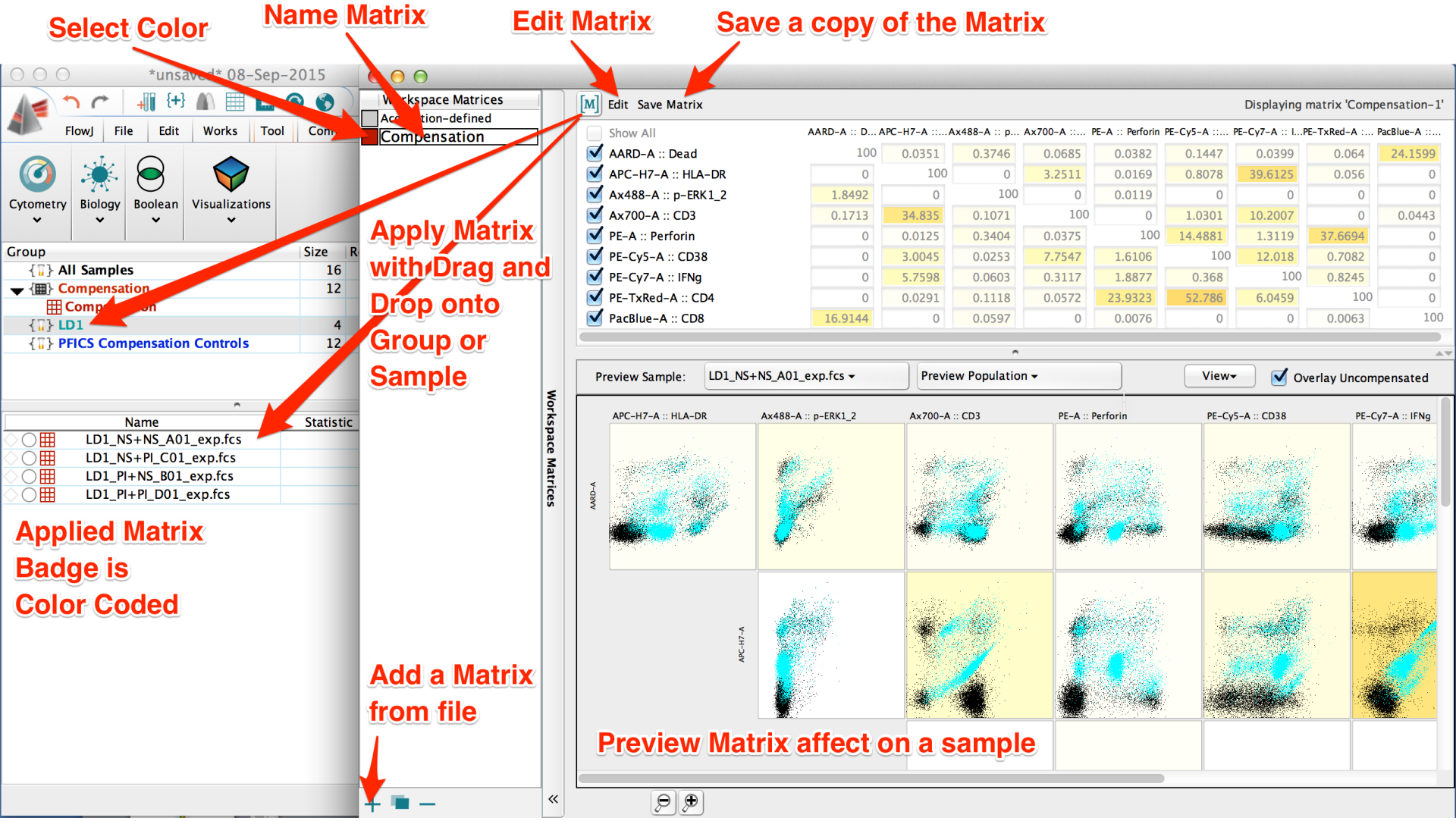
Use Positive drop down list

to Choose Positive population

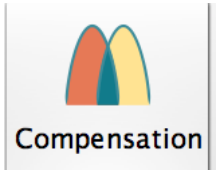


- Note that you can always create your own gates on a sample and then choose those from the drop down menus.
- When set up is complete, select View Matrix (top right) to Modify, Apply, Save or Preview the matrix you've created.

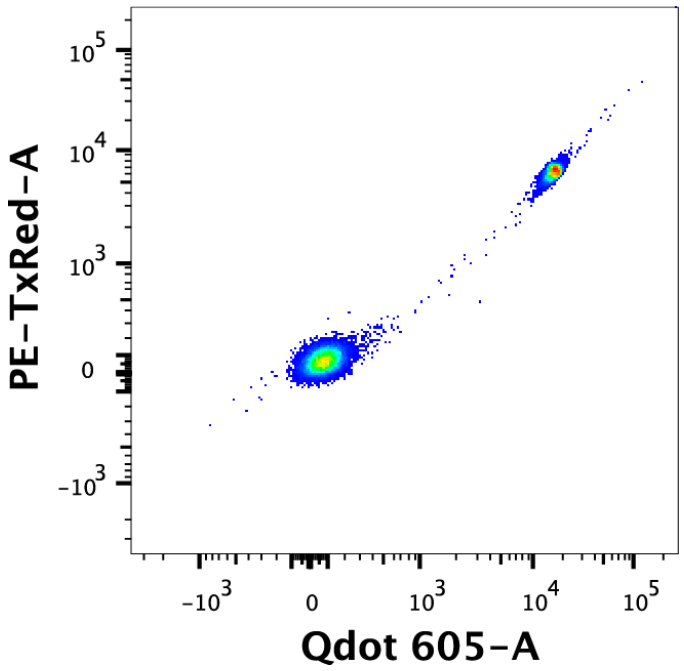
Compensation



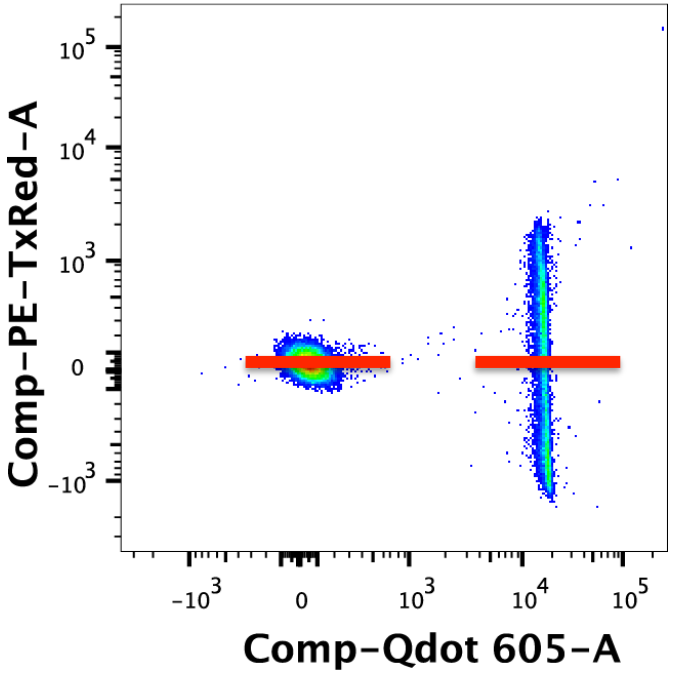
Effect of Compensation



Uncompensated

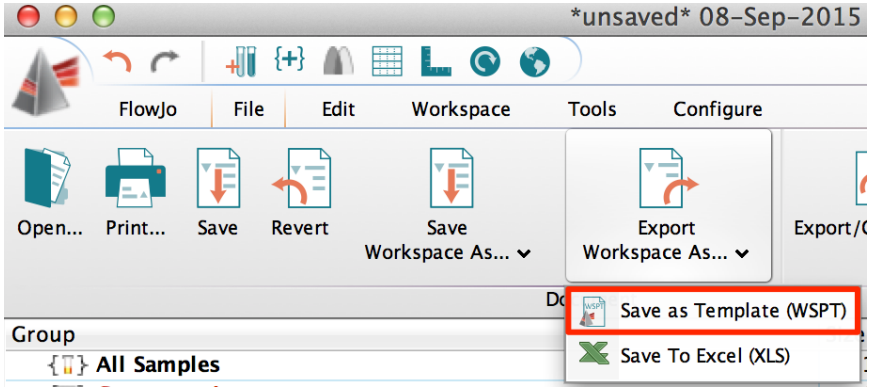


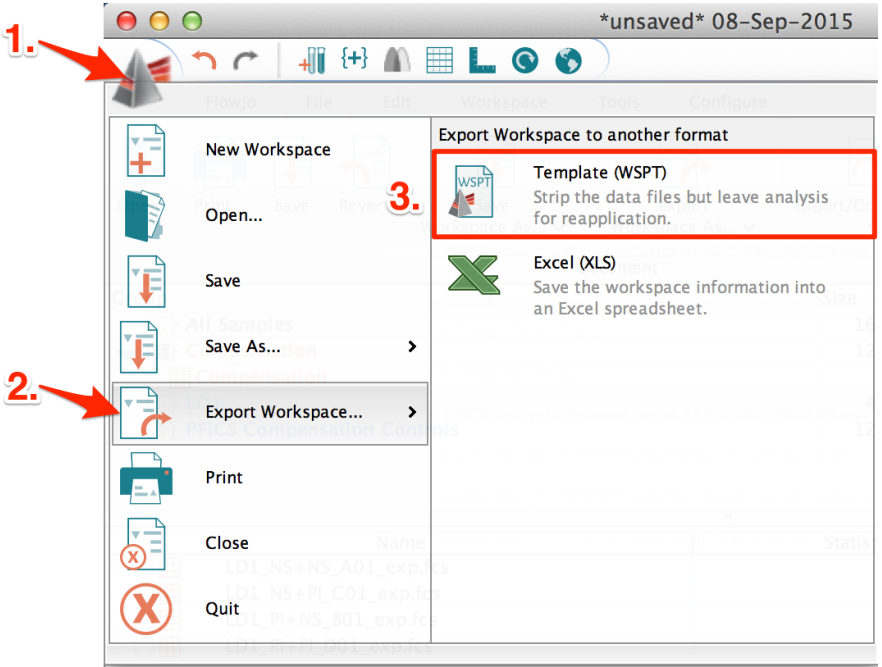
Compensated



Workspace Templates

- Allows saving all analysis reports in your workspace without data.
- Streamlines repetitive analysis of multiple runs using the same staining panel(s).
- File Tab → Document Band → Export Workspace As… Save as a Template

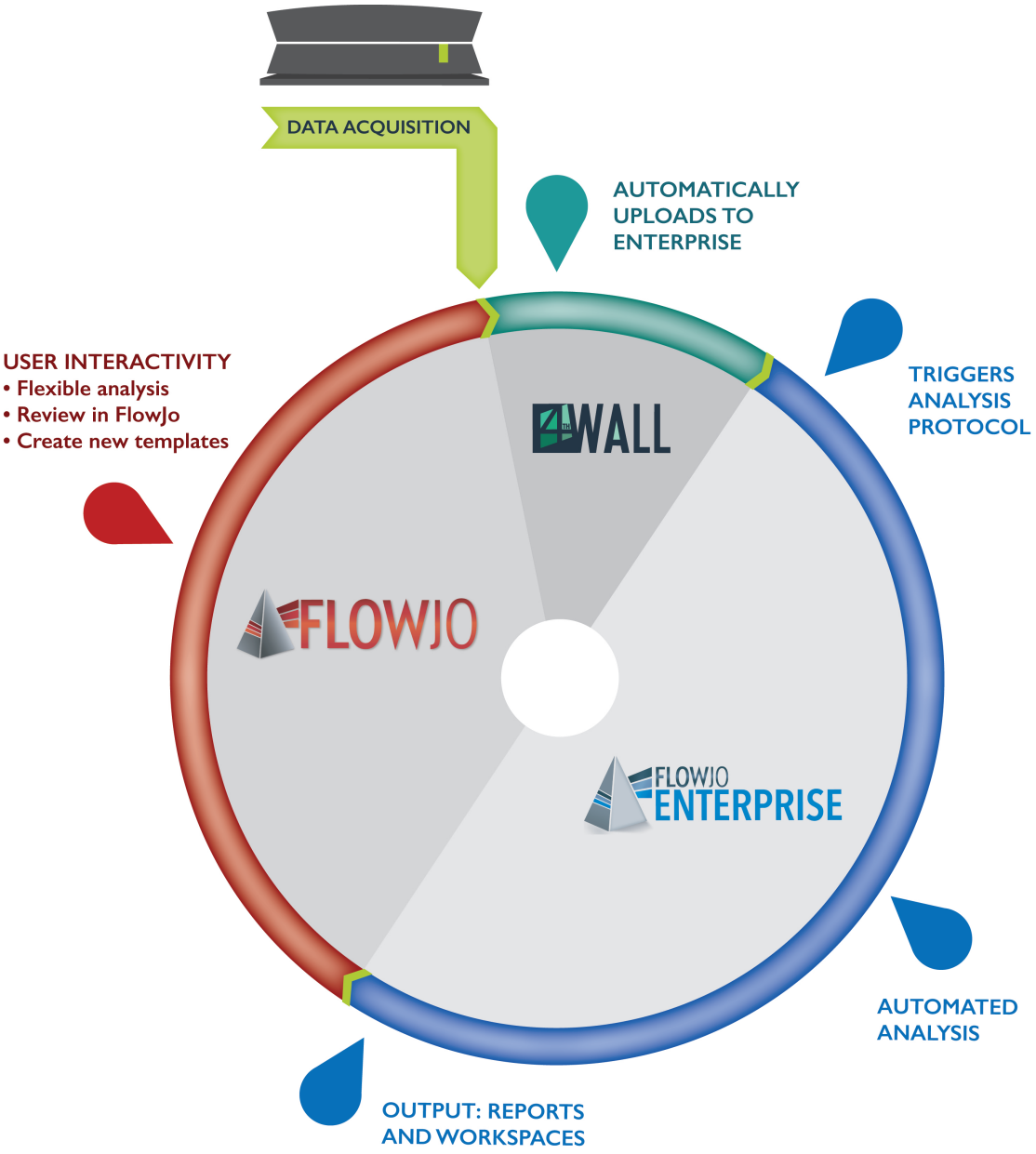




FlowJo Enterprise:

- is a server-based version of FlowJo v10, designed to assist with data archiving, analysis, and report generation for high dimension, high throughput flow or mass cytometry data.
- can handle data upload directly from the cytometer, store it on a secure server, and provide computational power and automated analysis features for scientists.
- is an optional add-on component of the FlowJo Licensing Server (FLS) institutional site license.
- is offered as 1 of 4 tiered packages, with each tier introducing additional features and levels of service.

FlowJo Enterprise Components



Email:
enterprise@flowjo.com
for information

The Plate Editor

- Viewer to add keywords in a plate format
- Located in the visualizations Band within the Tools Tab
- Add new keyword/value pairs to the right. Drag and drop on selected wells.

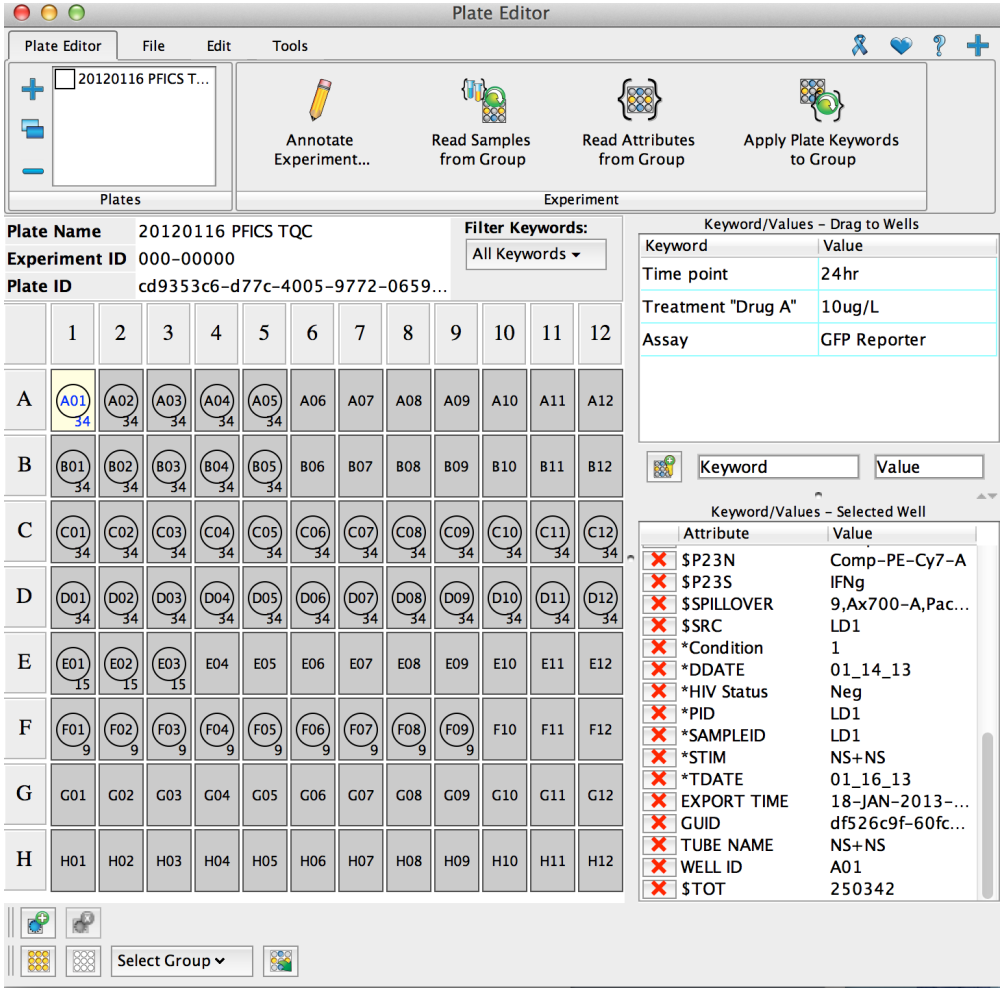
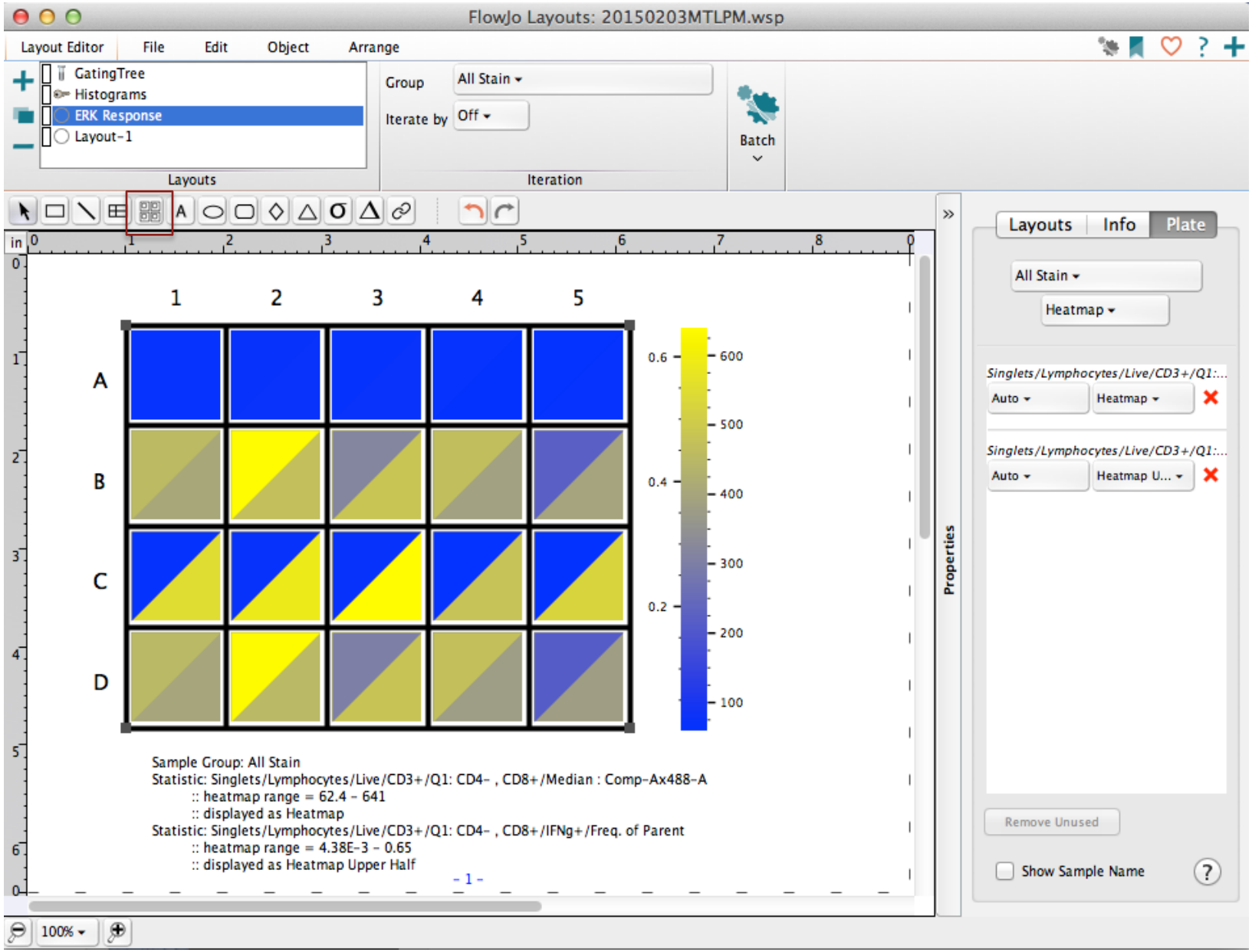


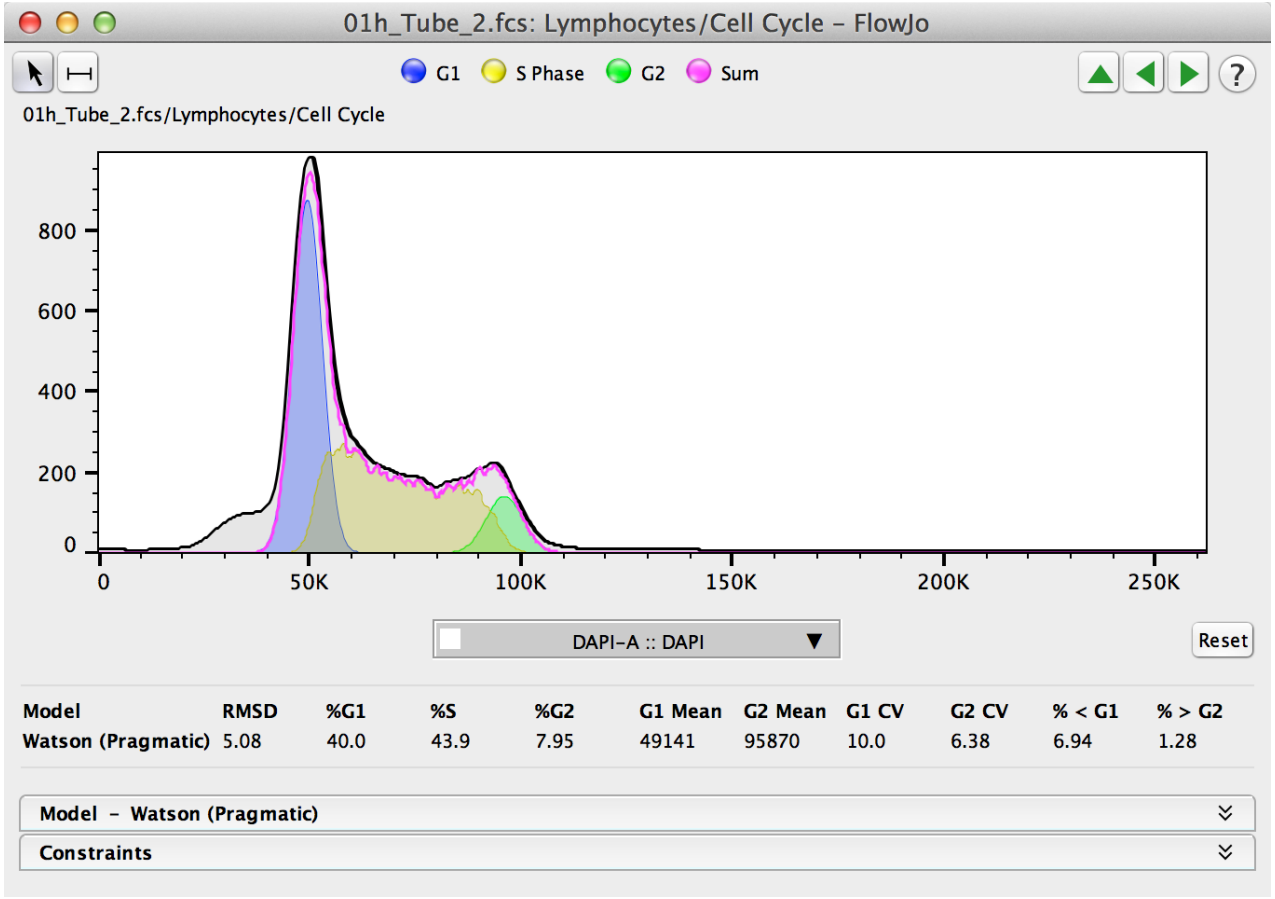
Plate Visualizations



Cell Cycle Analysis

- The Cell Cycle platform allows 1D modeling of cell cycle phases based on DNA content

- V10.1 has 1D Watson and Dean-Jett-Fox models.



Additional Training Resources

- Webinars on basic and advanced features of FlowJo, held on the 1st and 3rd Thursday of each month.
- Webinar Schedule can be found at http://www.flowjo.com/webinars/
- Technical Documentation for V10 can be found at http://docs.flowjo.com/
- The Daily Dongle provides tips, tricks and answers to common questions.

  http://flowjo.typepad.com/

Questions?

- FlowJo is here to help with all your cytometry analysis needs.
- Contact techsupport@flowjo.com for general questions and support.
- Contact timc@flowjo.com for science questions, additional training resources and information on FlowJo Enterprise.

Thank You!